

Research Article



Treatment of rotenone induced neurodegeneration by taurine and hesperidin

Eman S. Abdel-Reheim^{1*}, Basant M. Mahmoud², Hanan A. Soliman², Basma S. Ismail² ¹Physiology Branch, Zoology Department, Beni Suef University, Beni Suef, Egypt. ²Biochemistry Branch, Chemistry Department, Beni Suef University, Beni Suef, Egypt.

Received: 6/31/2017; Accepted: 7/28/2017 Available online: 1st August 2017

Abstract: Taurine (2-aminoethane sulphonic acid) is a sulphonic acid which is derived from cysteine and is widely distributed in animal tissues. It is one of the most abundant amino acid in mammals and plays several crucial roles including modulation of calcium signaling, osmoregulation and membrane stabilization. Hesperidin occurs in the cells in crystalline, feather-like aggregates or sphaerocrytalline masses and it exhibits pharmacological and biological properties such as anti-inflammatory, anticarcinogenic, inhibit bone loss, lowering of lipid, hypoglycaemic and antioxidant activities. The current study was performed to evaluate the effect of taurine and hesperidin on neurodegeneration resulted from rotenone administration by a dose of 1.5 mg/kg b.wt three times per week for two months. Also we summarize recent findings emphasizing the role of catecholamines neurotransmitters, Tyrosine hydroxylase and oxidative stress in neurodegenerative disease model. These rats received taurine and hesperidin through gastric intubation daily for one month after rotenone administration. The results revealed that taurine and hesperidin treatment significantly ameliorated the decreased levels of the catecholamines neurotransmitters and Tyrosine hydroxylase which were decreased as after rotenone injection. Moreover, taurine and hesperidin treatment ameliorated lipid peroxidation and catalase levels.

Keywords: Rotenone, Taurine, Hesperidin, Neurodegeneration, Catecholamines, Oxidative Stress.

Introduction

Taurine (2-amino-ethanesulfonic acid) is a β -amino acid and is the most abundant amino acid in mammals, being widely distributed in the central nervous system (CNS) occupying the second place after glutamate in relation of its concentration which differs depending on the brain activity regions and animal species studied and presenting different functions which have been studied for their potential in neurology as a trophic factor in brain development, in calcium transport regulation, in the eardrum integrity, as neuromodulator, neurotransmitter, osmoregulator and for its neuroprotective action (Wu and Prentice, 2010).

Hesperidin (Hsp) is a flavanone glycoside (flavonoid C28H34O15) found abundantly in citrus fruits (Walaa et al, 2014). Hesperidin occurs in the cells in crystalline, feather-like aggregates or sphaerocrytalline masses (Priya and Vijayalakshmi, 2016). Hsp, a flavonoid that is particularly abundant in oranges and lemons, exerts antioxidant, antihypertensive, anticarcinogenic, antiviral and anti-inflammatory effects (Parhiz et al, 2015). In addition, it can cross the blood-brain barrier (BBB) and can protect the neurons against various types of insult associated with neurodegenerative diseases, including Parkinson's disease (PD), Alzheimer's disease (AD) and Huntington's disease (HD) (Santa et al, 2016).

*Corresponding Author:

Dr. Eman Abdel-Reheim, Zoology Department Physiology branch, Beni Suef University, Beni Suef, Egypt. **E-mail:** adel men2020@yahoo.com Rotenone (ROT) is a lipophilic, naturally occurring compound and mainly derived from the roots and stems of *Lonchocarpus and Derris species*. ROT acts as a strong inhibitor of the mitochondrial respiratory chain (MRC) complex I (Sabrina *et al*, 2017).

The aim of this study was to evaluate the treatment effect of taurine and hesperidin against rotenone induced neurodegeneration.

Materials and Methods

Experimental animals:

White male albino rats (Rattus norvegicus) weighing between 100 g and 120 g were used as experimental animals in the present investigation. They were obtained from the animal house of National Research Institute, El-Giza, Egypt. They were kept under observation for about 10 days before the onset of the experiment to exclude any intercurrent infection. The chosen animals were housed in plastic cages with good aerated covers at normal atmospheric temperature $(25\pm5^{\circ}C)$ as well as under good ventilation and received water and standard balanced diet. All the procedures were performed in accordance with the Institutional Animal Ethics Committee in Beni-Suef University recommendations.



Chemicals:

Rotenone (ROT), Taurine (Tau), Hesperidin (Hsp), Dimethyl sulfoxide (DMSO) and Carboxy methyl cellulose (CMC) were purchased from Sigma Chemical Company, St. Louis, MO, USA. The dose selection for each compound was based on previously published studies.

Doses and treatment:

The dose of rotenone used in this study was 1.5 mg/kg.b.wt. dissolved in 1ml DMSO+ 4ml saline (Abdelkader *et al*, 2014). It was administered three times per week for two months by subcutaneous injection. Taurine and hesperidin were administered by gastric intubation in a dose of 100 mg / kg.b.wt. dissolved in distilled water (Ezekiel *et al*, 2015) and 50 mg/kg.b.wt. dissolved in 1% CMC (Balakrishnan and Menon, 2007) respectively, daily for one month after rotenone administration.

Experimental design:

The number of rats used in the present study is 36 rats. They were allocated into 6 groups; designed as follow:

Group 1: The control group, it was given the vehicle of the administered substances for three months.

Group 2: The taurine treated group.

Group 3: The hesperidin treated group.

Group 4: The rotenone injected group.

Group 5: The rotenone + taurine group, the rats of this group were administered with taurine daily for one month after two months of rotenone injection.

Group 6: The rotenone + hesperidin group, the rats of this group were administered with hesperidin daily for one month after two months of rotenone injection.

At the end of three months, six animals of each group were sacrificed under mild diethyl ether anesthesia. Brain from each animal was rapidly excised after dissection. 0.5g from brain was homogenized in 5ml 0.9% sterilized NaCl (10% w/v) using teflon homogenizer (Glas-Col, Terre Haute, USA) for biochemical analysis.

Biochemical analyses:

Epinephrine (E) and Norepinephrine (NE) contents were measured fluorometrically. Tyrosine hydroxylase (TH) was measured by using western blotting assay. In the midbrain, lipid peroxidation

was determined by the method of Ohkawa *et al.*, (1979) and catalase activity was determined by the method of Aebi, (1984).

Statistical analysis:

The Statistical Package for the Social Sciences (IBM SPSS for WINDOWS7, version 20; SPSS Inc, Chicago) was used for the statistical analysis of the results. Comparative analysis was conducted by using the general linear models procedure (IBM SPSS). p>0.05 were considered statistically non-significant, while p<0.05 were considered statistically significant.

Results

Data summarized in Table 1 and figures 1, 2, 3 and 4 indicating the effect of rotenone administration and treatment with taurine and hesperidin on midbrain epinephrine, norepinephrine and tyrosine hydroxylase in striatum. Our results revealed that the administration of rotenone produced marked impairment demonstrated by significant decrease in these parameters as compared to normal rats, while oral administration of taurine and hesperidin significantly increased these decreased levels of brain epinephrine, norepinephrine and tyrosine hydroxylase when compared with the rotenone administered rats recording noticeable significant amelioration as compared to the control ones.

Table 2 and figures 5 and 6 showing the effect of the tested taurine and hesperidin on the midbrain oxidative stress markers and antioxidant defense system in the midbrain of rotenone-administered rats. Lipid peroxidation (LPO) was significantly increased as a result of rotenone administration while treatment of rotenone-administered rats with taurine and hesperidin significantly decreased these elevated values and becomes near to those of control values. On the other hands, rats injected with rotenone exhibited a significant decrease in values of catalase activity as compared to control ones. While, the treatment with taurine and hesperidin produced a significant increase of this antioxidant enzyme activity as compared to the corresponding rotenone administered group pointing to a marked amelioration.

Table 1: Effect of taurine and hesperidin on epinephrine, norepinephrine and tyrosine hydroxylase levels in rotenone-administered rats.

Groups	E (μg)	NE (µg)	TH
Normal	41.50 ± 2.08 ^c	634.85 ± 4.87 ^d	1.03 ± 0.003°
Taurine	42.38 ± 1.04 ^c	649.85 ± 6.12^{d}	1.02 ± 0.01°
Hesperidin	42.70 ± 2.33°	639.10 ± 14.55^{d}	1.04 ± 0.01°
Rotenone	15.20 ± 0.53^{a}	212.25 ± 3.23ª	0.29 ± 0.02^{a}
Rotenone+Taurine	34.47 ± 1.18^{b}	511.87 ± 7.96°	0.85 ± 0.01^{b}
Rotenone+Hesperidin	$34.26 \pm 1.61^{\text{b}}$	468.07 ± 12.36 ^b	0.82 ± 0.03^{b}
LSD	2.00	13.00	0.029

*Values significantly different to control at $(p \le 0.05)$.

*Data are expressed as mean ± SE.

*Values which share the same superscript symbol are not significantly different.

*F-Probability: P < 0.05

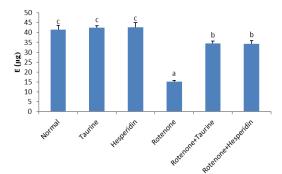


Fig. 1: The effect of taurine and hesperidin administration on midbrain epinephrine level in normal control and rotenone administered rats.

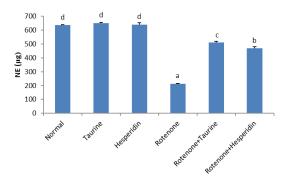


Fig. 2: The effect of taurine and hesperidin administration on midbrain norepinephrine level in normal control and rotenone administered rats.

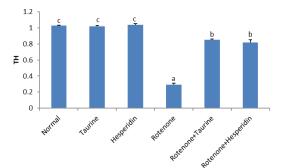


Fig. 3: The effect of taurine and hesperidin administration on striatum tyrosine hydroxylase level in normal control and rotenone administered rats.

Fig 4: Western blotting analysis showing the effect of taurine and hesperidin administration on striatum tyrosine hydroxylase levels in normal control and rotenone administered rats.

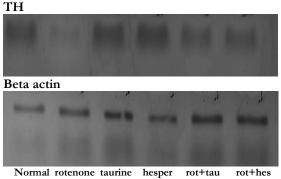


 Table 2: Effect of taurine and hesperidin on midbrain oxidative stress in normal control and rotenoneadministered rats.

Groups	MDA (nmol/g.tissue)	CAT
Groups	MDA (IIII01/g.ussue)	(U/g.tissue)
Normal	1.23 ± 0.06^{a}	126.85 ± 0.56^{d}
Taurine	1.75 ± 0.05^{a}	123.80 ± 1.24^{d}
Hesperidin	1.19 ± 0.04^{a}	124.95 ± 0.93^{d}
Rotenone	22.16 ± 1.08^{d}	61.22 ± 1.46^{a}
Rotenone+taurine	3.16 ± 0.12^{b}	$101.60 \pm 3.45^{\text{b}}$
Rotenone+Hesperidin	4.49 ± 0.21°	108.56 ± 1.95°
LSD	0.64	2.6

*Values significantly different to control at $(p \le 0.05)$.

*Data are expressed as mean ± SE.

*Values which share the same superscript symbol are not significantly different.

*F-Probability: P < 0.05

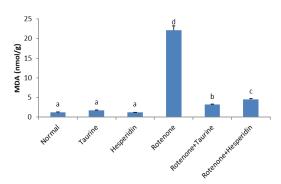


Fig. 5: The effect of taurine and hesperidin administration on midbrain malondialdehyde level in normal control and rotenone administered rats.

Fig. 6: The effect of taurine and hesperidin administration on midbrain catalase activity in normal control and rotenone administered rats.

Discussion

Mitochondrial dysfunction and neuroinflammation are common features of chronic neurodegenerative diseases of the central nervous system (CNS). Both conditions can lead to increased oxidative stress by excessive release of harmful reactive oxygen and nitrogen species, which further promote damage of neurons and subsequent inflammation resulting in a feed-forward loop of neurodegeneration (Roman and Olaf, 2015).

Rotenone (ROT), a potent retinoid which is used as a pesticide and insecticide, has been shown to cause systemic inhibition of mitochondrial complex I of activity, with consequent degeneration dopaminergic neurons within the substantia nigra and striatum, as observed in PD (Carriere et al, 2014). It is highly lipophilic in nature enabling it to cross the blood-brain barrier (BBB), and unlike many other toxic agents it can bypasses the dopamine transporter for cellular entry (Nidhika et al, 2016). Once in the cell, it accumulates in subcellular organelles including the mitochondria where it binds specifically to complex I leading to disruption of mitochondrial respiration and increasing the production of reactive oxygen species (ROS) and oxidative stress (Nistico et al, 2011).

Our results obtained in the present work can be discussed in two main aspects: first, the development of neurodegenerative disease model induced by rotenone; second, the effects of taurine and hesperidin against changes induced by rotenone. According to the first aspect, it was possible to observe that the injection of rotenone at the dose of 1.5mg/kg.b.wt. caused several changes antioxidant system that in can induce neurodegeneration. Our results showed that administration of rotenone caused a significant decrease in epinephrine, norepinephrine and tyrosine hydroxylase levels. These results run parallel to those of Nidhika et al., (2016) whose results showed that administration of rotenone for significantly 28 davs decreases levels of norepinephrine, serotonin and dopamine. Gunduluru and Wudavagiri (2014) speculated that norepinephrine levels were decreased in rotenoneinduced rats may be due to the 50% loss of neurons in Parkinson's disease. Although the rotenone microtubule-depolymerizing activity is not cell type-specific either, it significantly contributes to the selective dopaminergic neurons death because it disrupts microtubule-based transport of dopamine vesicles (Yong et al, 2005). These results in accumulation of vesicle in the soma, which leads to oxidative stress increasing due to oxidation of dopamine leaked from the vesicles (Eisenhofer et al, 2004). Thus, the combination of both rotenone activities would generate far more ROS in dopaminergic neurons than in other types

of cells, rendering these neurons vulnerable particularly to this environmental PD toxin. Thus, catecholaminergic neurons that depend on long range microtubule based transport to move their oxidizable cargoes are vulnerable to microtubuledepolymerizing agents such as colchicine or rotenone (Yong et al, 2005). In the same way, Lamberto et al., (2007) revealed that the concomitant decline in the levels of dopamine and norepinephrine mediators might indicate the inhibition of neurotransmitter synthesis through the tyrosine hydroxylase inactivation and/or mitochondrial dysfunction and cell damage. As for tyrosine hydroxylase level, Zhaoqiang et al., (2015) indicated that the tyrosine hydroxylaseimmunoreactive neurons number and the tyrosine hydroxylase expression levels were decreased in the substantia nigra of rats treated with rotenone. It has been found that treatment with rotenone leads to nigrostriatal dopaminergic neurodegeneration, resulting in a loss of tyrosine hydroxylase-positive neurons in the substantia nigra (Nuramatjan et al, 2016). In the same way, Amparo et al., (2013) revealed that 1-methyl-7-phenylpyridine (MPP+) or rotenone treatment decreased the number of tyrosine hydroxylase-positive neurons in chicken midbrain dopaminergic neuron culture, thus allowing the possibility to test neuroprotective capabilities of potential trophic factors in vitro.

In our study, administration of taurine and hesperidin to rotenone treated rats resulted in elevation of the lowered epinephrine, norepinephrine and tyrosine hydoxylase levels as compared to rotenone group. These results go parallel with Chepkova et al., (2005) whose results revealed that extracellular addition of taurine could activate the glycine receptor and the specific taurine transporter, increase the efflux of dopamine, following the release of other neurotransmitters in the central nervous system. Also, Xiaohui et al., (2013) showed that the number of synaptic vesicles was increased in the brains exposed to arsenic and taurine together, therefore these results indicated that taurine had a protective effect on the decreased of norepinephrine that might be a reasons for taurine protection effect on the arsenic decreased induced the neurotransmitters concentration in the brains. They also showed that the expression of tyrosine hydroxylase (TH), tryptophan hydroxylase (TPH) and dopamine beta hydroxylase (DBH) mRNA were significantly increased in the cerebellum of mice exposed to arsenic and taurine together, except that the changed in TPH mRNA in the cerebrum wasn't significant and hence these results indicated that taurine had a protective effect on the decreased levels of biogenic amine neurotransmitter in the brain of mice exposed to arsenic through elevating the DBH, TH, and TPH mRNA expression.

On the other hands, Priya *et al.*, (2014) stated that the protective effect of hesperidin may be due to neurotransmitters such as norepinephrine and epinephrine regulation which may play an important role in protection of neurobehavioral activities and antioxidant properties. Also, Priya and Vijayalakshmi (2016) observed that tyrosine immunoreactive neurons of striatum and mid brain were reduced after treatment with 6hydroxydopamine and this decrease was reversed by hesperidin administration and much better by hesperidin in combination with L-dopa.

The oxidative stress usually results from either excessive production of ROS, impaired antioxidant system, dysfunction of mitochondria or a combination of these factors (Małgorzata and Andrzej, 2016). Sushruta et al., (2012) revealed that decreased activities of antioxidant enzymes such as glutathione peroxidase, superoxide dismutase, catalase and glutathione in neurodegenerative states signify the role of antioxidant potential reduction during neurodegeneration. Rotenone inhibits complex I of the protein mitochondrial electron transport chain leading to reduced ATP production and leakage of electron that can form reactive oxygen species such as superoxide, subsequently causing glutathione levels reduction and oxidative stress (Johnson and Bobrovskaya, 2015).

In the present study, our results revealed that rotenone administration resulted in increased level of lipid peroxidation and caused significant decrease in catalase level. These results go parallel with Omar et al., (2015) who observed that rotenone resulted in increased nitric oxide and lipid peroxidation concentrations while decreasing antioxidant mechanisms such as catalase, reduced glutathione level and Paraoxonase 1(PON1) activities in different brain regions. In the same way, Neha and Pallavi (2012) observed that the rotenone per selenium group showed a significant increase in the levels of thiobarbituric acid (which is an indication of lipid peroxidation extent), decrease in the superoxide dismutase and gluathione reduced levels in the brain as compared to the vehicle treated control animals, therefore all these indicate an oxidative stress increase in the brain of rotenone treated animals. Also, Yasser et al., (2016) stated that rotenone induced a significant increase in striatal lipid peroxidation in the striatum. The depletion of glutathione reduced, ATP and the increase of malondialdehyde and protein carbonyl levels might support the oxidative stress occurrence resulting in peroxidation of lipid and protein and mitochondrial dysfunction, which lead to energy crisis and cell damage (Lamberto et al, 2007).

Administration of taurine and hesperidin to rotenone intoxicated rats resulted in a significant decrease in malondialdehyde level and resulted in a

significant increase in catalase level. These results are in accordance with (Zuhal and Nedret, 2011) who stated that the useful effects of taurine in biological systems as an antioxidant have been attributed to its ability to stabilize biomembranes, to scavenge ROS, and to decrease the peroxidation of unsaturated membrane lipids. It has been shown that taurine exhibits its antioxidant capacity by the antioxidant system enhancement, formation of chloramines with hypochlorous acid and replacement of glutathione in biological systems during oxidative stress (Devi and Anuradha, 2010). Jagadeesan and Sankarsami (2007) concluded that taurine reduces the oxidative stress through lipid peroxidation inhibition and also through increased catalase, glutathione peroxidase and superoxide dismutase which replenish glutathione reduced stores and allows for correct cell defense against ROS. Also, Ezekiel et al., (2015) stated that taurine may inhibit lipid peroxidation by superoxide dismutase and glutathione peroxidase induction and also observed that the antioxidant taurine prevented oxidative stress and cellular antioxidants loss and suggested that taurine protected forebrain from restrain stress-induce oxidative damage and also, shown that taurine offsets lipid peroxidation either through ROS scavenging directly or by binding to copper ion or ferrous ion through its sulphonic acid group. In the same way, Rosemberg et al., (2010) revealed that treatments with taurine prevented the alterations promoted in catalase and superoxide dismutase activities by ethanol (EtOH), suggesting a modulatory role in enzymatic antioxidant defenses. On the other hands, Hiam et al., (2012) revealed that there is an increase in the glutathione reduced content and decrease in the extent of lipid peroxidation with the hesperidin treatment and demonstrated that hesperidin administration to group treated with Chlorpyrifos (CPF) showed an improvement in the levels of endogenous antioxidant enzymes (superoxide dismutase, catalase and glutathione S transferase), glutathione reduced and acetylcholinesterase. In the same regard, Syed et al., (2011) demonstrated that significantly antioxidant enzymes, glutathione peroxidase, catalase, glutathione reductase and superoxide dismutase and content of glutathione depleted activity in Middle cerebral artery occlusion (MCAO) group were protected significantly in MCAO group pretreated with hesperidin. The properties antioxidant/neuroprotective of flavanoids including hesperidin, involves metal ions chelation such as copper and iron resulted in inhibition of transition metal-catalyzed free radical formation (Kuppusamy et al, 2013). Pretreatment significantly with hesperidin resulted in and nitrite malondialdehyde concentration improvements in striatum and cortex regions of the brain suggesting its antioxidant like effects, therefore hesperidin has been reported to offer neuroprotection by lipid peroxidation side chain termination rather than extracellular non-lipid radicals scavenging that initiate lipid peroxidation (Vaibhav *et al*, 2011). In the same way, hesperidin therapy could significantly attenuate formation of ROS by reducing the levels of thiobarbituric acid reactive substances (TBARS) and restored glutathione reduced and antioxidant enzyme activity to physiological levels in the brain (Kamisli *et al*, 2013) and cultured PC12 cells (Kuppusamy *et al*, 2013).

Conclusion

Taurine and hesperidin administration exhibited a beneficial therapeutic effect on rats with neurodegeneration due to their ability to regulate neurotransmitters and due to their antioxidative properties.

References

- Abdelkader NF., Safar MM. and Salem HA. Ursodeoxycholic Acid Ameliorates Apoptotic Cascade in the Rotenone Model of Parkinson's Disease: Modulation of Mitochondrial Perturbations. *Mol Neurobiol.* (2014); 53: 810-817.
- Aebi H. Catalase in vitro. Methods Enzymol. (1984); 105: 121-126.
- Amparo T., Xiaolai Z., Bjorn S. and Kerstin K. Establishment of a Survival and Toxic Cellular Model for Parkinson's Disease from Chicken Mesencephalon. *Neurotox Res.* 2013; 24(2): 119–129.
- Balakrishnan A. and Menon VP. Protective effect of hesperidin on nicotine induced toxicity in rats. *Indian Journal of Experimental Biology*. (2007); 45:194-202.
- Carriere CH., Kang NH. and Nies LP. Neuroprotection by Valproic acid in an intrastriatal rotenone model of Parkinson's disease. *Neuroscience*. (2014); 267: 114-121.
- Chepkova AN., Sergeeva OA. and Haas HL. Long-lasting enhancement of corticostriatal transmission by taurine: role of dopamine and acetylcholine. Cell *Mol Neurobiol.* (2005); 25: 767-776.
- Devi SL. and Anuradha CV. Oxidative and nitrosative stress in experimental rat liver fibrosis: protective effect of taurine. *Environmental Toxicology and Pharmacology*. (2010); 29(2): 104-110.
- Eisenhofer G., Kopin IJ. and Goldstein DS. Leaky catecholamine stores: undue waste or a stress response coping mechanism? *Ann. N.Y.Acad. Sci.* (2004); 1018: 224-230.
- Ezekiel I., Ayo JO., Tanko Y. and Magaji RA. Effect of Taurine on Acute Restraint Stress Induced Biochemical Alterations in Wistar Rats. International *Journal of Scientific* & Engineering Research. (2015); 6(5): 2229-5518.
- Gunduluru S. and Wudayagiri R. Protective role of Bacopa Monnieri on induced Parkinson's disease with particular reference to catecholamine system. *International Journal of Pharmacy and Pharmaceutical Sciences*. (2014); 6(7): 379-382.
- Hiam RA., Amira AM., Eman MS. and Kamal AF. Influence of Hesperidin combined with Sinemet on genetical and biochemical abnormalities in rats suffering from Parkinson's disease. *Life Sci J.* (2012); 9(4): 930-945.

- Jagadeesan G. and Sankarsami PS. Hepatoprotective effects of taurine against mercury induced toxicity in rats. *Journal of Environmental Biology*. (2007); 28(4): 753-756.
- Johnson ME. and Bobrovskaya L. An Update on The Rotenone Models of Parkinson's Disease: Their Ability to Reproduce the Features of Clinical Disease and Model Gene–Environment Interactions. *Neurotoxicology*. (2015); 46: 101–116.
- Kamisli S., Ciftci O., Kaya K., Cetin A., Kamisli O. and Ozcan C. Hesperidin protects brain and sciatic nerve tissues against cisplatin-induced oxidative, histological and electromyographical side effects in rats. *Toxicology and Industrial Health.* (2013); 31(9): 841-51.
- Kuppusamy T., Jagadhesan N., Udaiyappan J., Thamilarasan M. and Mustafa ME. Antioxidant and antiinflammatory potential of hesperidin against 1-methyl-4phenyl-1, 2, 3, 6-tetrahydropyridine-induced experimental Parkinson's disease in mice. *International journal of nutrition, pharmacology, neurological diseases.* (2013); 3(3): 294-302.
- Kuppusamy T., Nady B., Thamilarasan M., Musthafa ME., Nagarajan RP., Subburayan K., Arokyasamy JT., Subash S. and Gilles JG. Neuroprotective Effects of Hesperidin, a Plant Flavanone, on Rotenone-Induced Oxidative Stress and Apoptosis in a Cellular Model for Parkinson's Disease. Oxid Med Cell Longev. (2013); 2013: 102741.
- Lamberto R., Mohamed NM., Silvia M., Olga SL., Gregorio MS. and Frank H. Ozone Therapy: Clinical and Basic Evidence of Its Therapeutic Potential. *ARCMED*. (2007); D-07-00225.
- Malgorzata N. and Andrzej G. The Role of the Reactive Oxygen Species and Oxidative Stress in the Pathomechanism of the Age-Related Ocular Diseases and Other Pathologies of the Anterior and Posterior Eye Segments in Adults. Oxidative Medicine and Cellular Longevity. (2016); 2016: 3164734.
- Neha S. and Pallavi B. Effect of Cynodon dactylon on rotenone induced Parkinson's disease. Orient Pharm Exp Med. (2012); 12(3): 167-175.
- Nidhika S., Sumit J. and Puneet K. Beneficial effect of antidepressants against rotenone induced parkinsonism like symptoms in rats. *Pathophysiology*. (2016); 23(2): 123-34.
- Nistico R., Mehdawy B., Piccirilli S. and Mercuri N. Paraquat- and rotenone-induced models of Parkinson's disease. *Int.J. Immunopathol. Pharmacol.* (2011); 24(2): 313-322.
- Nuramatjan A., Deyong L., Rutong R., Yilixiati X., Xiang M., Xin Z., Yi S., Hui L., Jiamin X., Yingcong M., Xianrong Q., Min Y., Feng X., Hongbin H. and Xiaoping P. Neuroprotective Effects of a Standardized Flavonoid Extract from Safflower against a Rotenone-Induced Rat Model of Parkinson's Disease. *Molecules*. (2016); 21(9): 1107.
- 23. Ohkawa H., Ohishi W. and Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal. Biochem.* (1979); 95: 351-8.
- Omar ME., Eman RY., Yasser AK., Nadia AM., Rehab FA., Enayat AO. and Amany AS. The effect of cannabis on oxidative stress and neurodegeneration induced by intrastriatal rotenone injection in rats. *Comparative Clinical Pathology*. (2015); 24(2): 359-378.
- 25. Parhiz H., Roohbakhsh A., Soltani F., Rezaee R. and Iranshahi M. Antioxidant and anti-inflammatory properties of the Citrus flavonoids hesperidin and hesperetin: An

updated review of their molecular mechanisms and experimental models. *Phytother Res.* (2015); 29: 323–331.

- Priya N. and Vijayalakshmi K. Anti-Parkinson effect of hesperidin in combinatin with L-DOPA on 6-OHDA induced Parkinsonism in wistar rats: a neurochemical, histopathological and immunohistochemical analysis. International *Journal of Pharm Tech Research*. (2016); 9(4): 266-273.
- Priya N., Vijayalakshmi K. and Khadira S. Investigation on the neuroprotective effects of hesperidin on behavioural activities in 6-OHDA induced parkinson model. *Int J Pharm Bio Sci.* (2014); 5(4): (B) 570 - 577.
- Roman F. and Olaf M. Interrelation of Oxidative Stress and Inflammation in Neurodegenerative Disease: Role of TNF. Oxidative Medicine and Cellular Longevity. (2015); 2015: 610813.
- Rosemberg DB., da Rocha RF., Rico EP., Zanotto-Filho A., Dias RD., Bogo MR., Bonan CD., Moreira JC., Klamt F. and Souza DO. Taurine prevents enhancement of acetylcholinesterase activity induced by acute ethanol exposure and decreases the level of markers of oxidative stress in zebrafish brain. <u>Neuroscience.</u> (2010); 171(3): 683– 692.
- Sabrina H., Alexius F., Bettina L., Ludwig S., Gabriele S. and Heidrun E. Mechanistic Investigations of the Mitochondrial Complex I Inhibitor Rotenone in the Context of Pharmacological and Safety Evaluation. Scientific Reports. (2017); 7: 45465.
- Santa C., Nadia F., Giovanni EL., Elvira V., Sebastiano G., Gioacchino C. and Michele N. Neurodegenerative Diseases: Might Citrus Flavonoids Play a Protective Role? *Molecules.* (2016); 21(10): 1312.
- Sushruta K., Hemant K., In SK. and Dong-Kug C. Reactive oxygen species and inhibitors of inflammatory enzymes, NADPH oxidase and iNOS in experimental models of Parkinson's disease. *Mediators of Inflammation*. (2012); 2012: 823902.
- Syed SR., Mohd MK., Ajmal A., Mohammad A., Gulrana K., Rizwana T., Hayate J., Mohammad SS. and Mohammed MS. Hesperidin ameliorates functional and histological outcome and reduces neuroinflammation in experimental stroke. <u>Brain Research. (2011)</u>; 1420: 93–105.

- Vaibhav G., Aditi A. and Anil K. Possible nitric oxide mechanism in the protective effect of hesperidin against ischemic reperfusion cerebral injury in rats. *Indian J Exp Biol.* (2011); 49: 609-618.
- Walaa GH., Howida SA. and Susan A. Protective Effects of Ruitn and / or Hesperidin Against Doxorubicin-Induced Hepatotoxicity. *International Journal of Clinical Nutrition.* (2014); 2 (1): 11-17.
- 36. Wu JY. and Prentice H. Role of taurine in the central nervous system. J. Biomed. Sci. (2010); 17: S1.
- Xiaohui L., Fengyuan P. and Yachen L. Protective Effect of Taurine on the Decreased Biogenic Amine Neurotransmitter Levels in the Brain of Mice Exposed to Arsenic. Adv Exp Med Biol. (2013); 776: 277-87.
- Yasser AK., Iman MM., Haitham SM., Neveen AN. and Heba SA. A study on the possible therapeutic role of Panax ginseng extract against a rat model of Parkinson's disease induced by intrastriatal rotenone injection. *Int. J. Clin. Exp. Med.* (2016); 9(2): 3831-3841.
- Yong R., Wenhua L., Houbo J., Qian J. and Jian F. Selective Vulnerability of Dopaminergic Neurons to Microtubule Depolymerization. *The journal of biological chemistry*. (2005); 280(40): 34105–12.
- Zhaoqiang Z., Xixun D., Huamin X., Junxia X. and Hong J. Lesion of medullary catecholaminergic neurons is associated with cardiovascular dysfunction in rotenoneinduced Parkinson's disease rats. *European Journal of Neuroscience*. (2015); 42(6): 2346–2355.
- Zuhal Y. and Nedret K. Effects of Taurine and Age on Cerebellum Antioxidant Status and Oxidative Stress. International Journal of Gerontology. (2011); 5(3): 166-170.

Cite this article as:

Eman S. Abdel-Reheim, Basant M. Mahmoud, Hanan A. Soliman and Basma S. Ismail. Treatment of rotenone induced neurodegeneration by taurine and hesperidin. *International Journal of Bioassays 6.8 (2017) pp. 5452-5458*.

DOI: <u>http://dx.doi.org/10.21746/ijbio.2017.08.002</u>

Source of support: Nil. Conflict of interest: None Declared