



PRELIMINARY PHYTOCHEMICAL SCREENING OF METHANOLIC–AQUA EXTRACT OF ACANTHOSPERMUM AUSTRALE LEAVES

Anthony Swamy Thangiah^{1*}, Mutuku Chrispus Ngule² and Obey Jackie K³

^{1* & 2}Department of Chemistry, University of Eastern Africa Baraton, P.O Box 2500-30100, Eldoret, Kenya

³Department of Medical Laboratory Sciences, University of Eastern Africa, Baraton Eldoret, Kenya

Received for publication: August 21, 2013; Accepted: September 11, 2013

Abstract: The study was carried to analyze the phytochemical constituents of the *Acanthospermum australe*. The plant samples were extracted using methanol and water in the ratio of 9:1. From the study the extract of *Acanthospermum australe* was found to contain tannins, saponins, terpenoids, phenols, alkaloids, steroidal rings but steroids and flavonoids were found to be absent. The presence of these important phytochemicals in the plant roots is a scientific justification of the plant use in the traditional treatment against various diseases affecting humans and animals. However, more research needs to be done to identify the specific compounds, their structural formulas and contribution towards medicinal values of the plant.

Keywords: Phytochemicals, pharmacological, leaves, *Acanthospermum*.

INTRODUCTION

Research on medicinal plants is of great importance taking in to account the old and new problems emerging day by day. Medicinal plants are available in nature and the grand's have information about their medicinal value traditionally. The information about natural healing methods was passed from grand's to children and grandchildren from generation to generation. With growing knowledge on technology and civilization this information transfer is no longer taken seriously in the society hence endangering the knowledge on traditional methods of treatment with one of them being the use of medicinal plants. This calls for a great need to have the knowledge on medicinal plants reserved and kept for future reference [1].

With increase in diseases caused by the modes of leaving and emerging drug resistant microbes, back-to-nature is becoming a common acronym to many people in the world today. The use of plants in the past tells clearly the fascinating relationship between mankind and plants since ancient times. Due to lack of clear knowledge on the mode of treatment of certain plants, people in the past have attributed the healing of diseases using medicinal herbs to supernatural forces due to their indisputable healing capability [2].

After many years of using medicinal herbs by humankind the isolation of active compounds such as morphine, quinine and alkaloids 200 years ago ushered in the dawn of a new era in the use of medicinal plants and marked the beginning of modern research in the use of plants to cure diseases [3]. According to Ameyaw [3], many plants in Africa such as *Cassia siame* (Lam), *Nauclea latifolia* (Benth), *Cryptolepis*

sanguinolenta (Lindl), *Azadiracha indica* and *Jatropha gossypifolia* have been tested for antimalarial properties. Many of the traditionally used plants in the African continent tested have shown great potential in biological activity [4].

Out of the 600 species of medicinal plants from 125 families tested against *P. gallinaceum* in chicks, *p.cathemerium* and *P. lophurae* in duckling 33 species were found to possess high potential in the treatment against the microbes, with the highest been plants *amaryllidaceae* and *simaroubaceae* family [5]. Medicinal plants have since ancient times been used to treat many illnesses which affect humankind even today. Many traditionalists have done this for quite some time and therefore prevented many deaths in the past few decades. However, this has been done with little scientific proof on the efficiency and the effects of the extracts on the affected individuals. Herbal medicine is still a matter of argument in the current world with many still doubting its efficiency. This has been due to greedy practitioners who want to become wealthy by pretending to know much about the diseases which their clients claim to have, hence leading to the application of wrong treatment and administration of wrong drugs which do not cure the patient and therefore leading to worsening of the situation or even death of the victim [6]. Much scientific data needs to be provided in order to create the needed confidence in the use of medicinal plants.

Due to their composition plants have been known to possess multiple medicinal properties hence enabling them to have several uses in the pharmaceutical industry. Studies on several plants have

Corresponding Author:

Dr. Anthony Swamy T,
Faculty of Chemistry,
University of Eastern Africa,
Baraton, P.O Box 2500-30100,
Eldoret, Kenya.



been done all over the world and plants have shown great potential in the treatment of diseases affecting both humans and animals. Study report on *Potentilla fulgens* has shown the plant to have anti-hyperglycemic, hypoglycemic, anti-hyperlipidemic, antitumor, antioxidant, anti-inflammatory and anti-ulcerogenic properties [7]. The use of medicinal plants is as old as man [8]. In the past few decades medicinal plants have been tested extensively and found to have several pharmacological uses such as, antibacterial activity, antifungal activity, anti-diabetic activity, anticancer activity, antioxidant activity, hepatoprotective activity, haemolytic activity, anti-inflammatory activity, larvicidal activity, anthelmintic activity, central nervous system activity and pain relief activity [9 & 10]. Many side effects associated with allopathic medicines and dependencies are common reasons why many people are hospitalized today. In order to counteract the effects, many people are now turning to nature in pure form to prevent and cure diseases using natural medicinal herbs or natural health alternatives [11].

MATERIALS AND METHODS

Sample Collection and Preparation:

The herb was randomly collected in the natural forest around University of Eastern Africa, Baraton. The plant samples were collected and identified by a taxonomist in the Biology Department, Baraton University. The samples were thoroughly mixed and spread to dry at room temperature in the chemistry laboratory for about three weeks. They were then ground into fine powder and put in transparent polythene bags.

Extraction procedure:

Using electric analytical beam balance 50 grams of the powdered leaves of the *Acanthospermum australe* was placed in 1000 ml conical flask, methanol and water were then added in the ratio of 9:1 respectively until the leaves were completely submerged in the solvent. The mixture was then agitated for thorough mixing. The mixture was kept for 24 hours on a shaker for effective extraction of the plant components. The extract was filtered using Butchner funnel; Whatman no.1 filter paper and a vacuum and pressure pump. The filtrate was re-filtered again using the same apparatus. The solvent was evaporated using rotary vacuum evaporator (R-11) with a water bath at 40°C. The extract was brought to dryness using vacuum and pressure pump at room temperature. The residue was then obtained and used for the experiment. The extract was then analyzed for the presence of phytochemicals.

Qualitative phytoconstituents analysis:

The extracts phytoconstituents analysis was done using standard procedures with slight modifications [12, 13 & 14].

1. Tannins:

About 0.5 g of the sample was put in a test tube and 20 ml of distilled water was added and heated to boiling. The mixture was then filtered and 1 % of FeCl₃ was added to the filtrate and observations made. Brownish green coloration indicated the presence of tannins.

2. Saponins:

The crude extract was mixed with 5 ml of water and vigorously shaken. The formation of stable foam indicated the presence of saponins.

3. Flavonoids:

A portion of the aqueous extract was added in a test tube. To this, 5 ml of dilute ammonia and 2 ml of concentrated sulphuric acid were added. The appearance of a yellow color indicated the presence of flavonoids.

4. Terpenoids:

The extracts of the plant material was taken in a clean test tube, 2 ml of chloroform was added and vigorously shaken, then evaporated to dryness. To this, 2 ml of concentrated sulphuric acid was added and heated for about 2 minutes. A greyish color indicated the presence of terpenoids.

5. Glycosides:

Salkowsks' test: The extract of the plant material was mixed with 2 ml of chloroform and 2 ml of concentrated sulphuric acid was carefully added and shaken gently, then the observations were made. A red brown colour indicated the presence of steroidal ring (glycone portion of glycoside).

6. Alkaloids:

The crude extract was mixed with 1% of HCl in a test tube. The test tube was then heated gently and filtered. To the filtrate a few drops of Mayer's and Wagner's reagents were added to the test tube. A resulting precipitate confirmed the presence of alkaloids.

7. Steroids.

Liebermann Burchard reaction: About 2 g of the extract was put in a test tube and 10 ml of chloroform added and filtered, 2 ml of the filtrate was mixed with 2 ml of a mixture of acetic acid and concentrated sulphuric acid was added along the side of the test tube. Blue green ring indicated the presence of steroids.

8. Phenols:

The plants extract was put in a test tube and treated with a few drops of 2% of FeCl_3 , blue green or black coloration indicated the presence of phenols.

RESULTS AND DISCUSSION

Table.1: Phytochemical study of *Acanthospermum australe*

Phytochemical	Observation	Inferences
Tannins	Blue-black	Present
Saponins	Stable foam	Present
Terpenoids	Grayish color	Present
Flavonoids	No yellow color	Absent
Phenols	Black coloration	Present
Alkaloids	Precipitate	Present
Steroids	No blue-green	Absent
Steroidal rings	Red-brown color	Present

From the study the extract of *Acanthospermum australe* was found to contain tannins, saponins, terpenoids, phenols, alkaloids, steroidal rings (glycone portion of the glycoside) but steroids and flavonoids were found to be absent (Table. 1). Alkaloids which are secondary metabolites, they can be defined as a cyclic compounds which have nitrogen in a negative oxidation state. They affect the chemical transmitters' action of the nervous system. They also have other pharmacological activities such as analgesic, antispasmodic, antihypertensive effects and anti-arrhythmic effects and antibacterial. Cryptolepine a major alkaloid in *S.acuta* was found to be an antimalarial agent [15]. Cryptolepine has also being used clinically to treat malaria, colic and stomach ulcers [16], and also used in anticancer drugs. According to Karou [17], much study has being done on pharmacological properties of alkaloids and proved to have antiprotozoal, cytotoxic and anti-inflammatory properties.

The presence of alkaloids in the plant justifies its' medicinal value. Alkaloids have been isolated from different plants and their medicinal values tested. The most important use of alkaloids already known with its originality from plants is the use of alkaloids compounds in the treatment of malaria. According to Ameyawn and Duker-Eshon [3], many of the antimalarial drugs used today are quinoline derivatives manipulated from cinchona species bark [18]. Alkaloids have been identified for their functions which include analgesic, antiplasmodic and antibacterial activity [19]. According to Ayitey [20], bitter leaves containing alkaloids are capable of reducing headache associated with hypertension.

The presence of Saponins shows the potential of the plants to be used to produce mild detergents and intracellular histochemistry staining to allow antibody

access to intercellular proteins [21]. They have been found to treat hypercholesterolemia, hyperglycemia, antioxidant, anti-inflammatory, central nervous system activities, anticancer and weight loss [21]. They are used to stop bleeding, treating wounds and ulcers as it helps red blood cells to precipitate and coagulate [22]. This can be attributed to ability of saponins to bind with glucose and cholesterol molecules. Saponins have also being associated with inhibitory effect on inflammation [22].

Saponins are used by the folkloric remedies of Kashmir (India) in treating wounds [23], this is because of their ability to cause red blood cells coagulation and therefore help in blood clotting, treating wounds and enteric ulcers problems [24]. Saponins have also been used to prevent hypercholesterolemia, antibiotic activity, anti-inflammatory and anti-diabetic.

Flavonoids are used as antioxidants because of their ability to scavage free radicals such as peroxide and hydroperoxide of lipid hydroxyl hence inhibiting oxidation that lead to degenerative diseases [25]. They can be used as anti-diabetic. According to Namki [26], flavonoids can be used to prevent synthesis of off flavours that are caused by fat oxidation. Flavonoids have been found to have antibacterial activity due to their ability to complex with extracellular and soluble proteins and to complex with bacterial cell wall [27]. Flavonoids are produced by plant in response to microbial infection and studies have shown that they have antibacterial activity against a wide range of micro-organisms [28].

Flavonoids are secondary metabolites with polyphenolic structure and synthesized in plants, through polypropanoid pathway [29]. Flavonoids have being classified in to six sub-groups which include flavones, flavanol, flavanone, flava-3-ols, isoflavone and anthocyanidin. Flavonoids are known to contain specific compounds called antioxidants which protect human, animal and plant cells against the damaging effects of free radicals. Imbalance between free radicals and antioxidants leads to oxidative stress which has being associated with inflammation, autoimmune diseases, cataract, cancer, Parkinson's disease, aging and arteriosclerosis. It also plays a role in heart diseases and neurodegenerative diseases. Flavonoids have also vasodilator activity a property which is useful in improving blood circulation in brain and in Alzheimer disease [30]. Leaf extract of *Ginkgo biloba* which contains flavonoids was used for improving blood circulation in brain varix. Several isoflavone can be used to improve blood circulation. Furanocoumarins can alter hexobarbital induced sleeping time, showed cytotoxic action and inhibited growth of tumor in mice. Free radicals including the hydroxyl, hydrogen peroxide, superoxide and lipid peroxide have being

associated with a number of diseases such as cardiovascular disease, cataracts, diabetes, gastrointestinal inflammatory diseases, cancer, asthma, liver disease, macular degeneration, periodontal disease and other inflammatory processes. These oxidants are produced during normal body chemical processes. They can be damaged through free-radical damage. Flavonoids such as quercetin, rosin, catechin and its derivatives and the oligomeric proanthocyanidins (OPCS) have shown in vitro studies to inhibit the oxidation of low-density lipoproteins (LDL).

Tannins are also secondary metabolites in plants. They are glycosides of gallic or protocatechic acids. Their astringent property makes them useful in preventing diarrhea and controlling hemorrhage due to their ability to precipitate proteins, mucus and constrict blood vessels [6]. This is the reason why traditional healers use plants rich in tannins to treat wounds and burns since they are able to cause blood clotting. Some tannins have been reported to inhibit HIV replication selectively besides the use of diuretics [31]. This shows how traditional medicinal plants rich in tannins can be used to control this dangerous disease. Tannins have also shown antiparasitic effects [32]. According to Bajal [33], tannins can also be used to protect the kidney since when taken the poliovirus, herpes complex virus and various enteric viruses are inactivated. Foods rich in tannins can be used to treat hereditary hemochromatosis which is a hereditary disease characterized by excessive absorption of dietary iron. According to Chung [34], many tannin molecules have been shown to reduce the mutagenic activity of a number of mutagens. The anti-carcinogenic and anti-mutagenic potentials of tannins may be related to their antioxidative property which is important in protecting cellular oxidative damage including lipid peroxidation. The growths of many fungi, yeast, bacteria and viruses have been proven to be inhibited by tannins. Tannins have also been reported to exert physiological effects, such as to accelerate blood pressure, decrease the serum lipid level, and produce liver necrosis and modulate immune responses. The dosage and kind of tannins are critical to these effects [34].

Glycosides another type of secondary metabolites are organic compounds from plants or animal sources in which a sugar is bound to a non-carbohydrate moiety. The term Glycoside is a collective term used for compounds formed with a glycosidic bonding between a sugar and another compound other than sugar. Cardiac glycosides have been used traditionally as arrow poisons or as heart drugs. They are used to strengthen the heart and make it function properly under controlled therapeutic dose. Cardiac glycosides bind to and inhibit Na^+/K^+ -ATPase, inhibition of Na^+/K^+ -

ATPase raises the level of sodium ions in cardiac myocytes, which leads to an increase in the level of calcium ions and an increase in cardiac contraction force [35]. The unexpected results relating cardiac glycosides with anticancer properties have created a great interest in this secondary metabolite. This has led to clinical trial of cardiac glycosides based drugs in clinics [36].

Terpenoids have medicinal value such as anti-carcinogenic, antimalarial, antimicrobial and diuretic activity [37 & 38]. Evaluation of the anti-inflammatory activity of three different Copaiba oleoresins showed that the crude extract of the plant and its' fractions of hexane, dichloromethane and methanolic extracts of *C. cearensis*, *C. reticulata* and *C. multijuga* have anti-inflammatory potential [39]. Terpenoids have also shown a great potential in treatment against disease causing microorganisms. Terpenoids have exhibited antibacterial activity against *E. coli*, *Staphylococcus*, *Pseudomonas aeruginosa* [40], *Proteus mirabilis* [40], *Klebsiella pneumoniae* [40 & 41], methicillin-resistant *S. aureus*, *Staphylococcus epidermidis* [41], *Listeria monocytogenes* [42], *Enterobacter cloacae*, yeast *Candida albicans* and fungi, *Aspergillus flavus* [43]. Terpenoids have also been found to reduce the growth of melanoma cells on mice after oral administration [44]. Terpenoids have been proved scientifically to kill mosquito larvae. Terpenoids extracted from *C. reticulata* species have shown potential in killing *A. aegypti* [45].

CONCLUSION

The presence of the important phytochemicals in the plant leaves is a scientific justification of the use of the plant in the traditional treatment against various diseases affecting humans and animals. The phytochemicals found in the plant have been analysed in other studies and found to have great pharmacological values. More research needs to be done to analyse the specific compounds in *Acanthospermum australe* plant, their structural make up, mode of action and their effect in the in vivo environment.

ACKNOWLEDGEMENTS

The authors of this paper are very much thankful to the Department of Chemistry, University of Eastern Africa; Baraton. The authors also thank the lab assistants for their dedication to ensure the smooth running of this research work.

REFERENCES

1. Prajapati DN, and Purohit SS (2003). *Agro's color atlas of medicinal plants*. Agrobios India: New Delhi.
2. Dodelis NI, Ferguson, James D, and Rattray D. *Magic and medicine of plants*. Pleasantville: New York, 1986

3. Ameyaw Y and Duker-Eshun G, The alkaloid content of the ethano-plant organs of three antimalarial medicinal plants species in the eastern region of Ghana. *Int.J.Chem*, 2009, 7(1):48-58.
4. Gbeassor M, Kossou Y, Amegbo K, Desouza C, Koumaglo K and Denke A, Antimalaria effects of eight African medicinal plants. *J. Ethanopharmacol*, 1963; 25(1):115-118.
5. O'Neil MJ, Bray DH, Boardman P, Phillipson JO, Warhurst DC. Plants as sources of antimalarial drugs.Part-1-in vitro test for evaluation of crude extracts form plants. *Plant Madica, N.V*, 1985 394-398.
6. Kokwaro JO, *Medicinal plants of east Africa*. Nairobi: University Press, 2009.
7. Koul K, Jaitak V and Kaul VK. Review on pharmaceutical properties and conservation measures of *Potentilla fulgens* Wall. ex hook. A medicinal endangered herb of higher Himalaya. *Indian Journal of Natural Products and resources*, 2011; 2(3):298-306.
8. Anthony ST, Ngule CM, Ngule ME, and Ramesh F. Qualitative analysis of phytoconstituents in *Tragia brevipes* plant. *International Journal of Pharmaceutical Research and analysis*, 2013; 3(2): 93-98.
9. Mir AM, Sawhney SS, and Jassal MMS. Qualitative and quantitative analysis of phytochemicals of *Taraxacum officinale*. *Wudpecker Journal of Pharmacy and Pharmacology*, 2013; 2(1): 01-05.
10. Sukirtha K and Growther L. Antibacterial, analysis of selected medicinal plants. *Journal of natural product and plant resource*, 2012; 2(6): 644-648.
11. Deshpande DJ. *A handbook of medicinal herbs*. Agrobios, Joghpur, India, 2010.
12. Trease G E & Evans WC. *Pharmacognosy*, 11th end, brailliere tindall, London, 1989, NV 45-50.
13. Harbome JB. *Phytochemical methods*. Chapman and hall ltd, London, 1973.
14. Sofowara A. *Medicinal plants and traditional medicine in Africa*. Spectrum books ltd, Ibadan Nigeria, 1993, 191-289.
15. Banzouzi JT, Prado R, Menan H, Valentin A, Roumestan C, Mallie M, Pelissier Y, and Blanche Y. Studies on medicinal plants of Ivory Coast: *Investigation of an active constituent phytomed*, 2004, 11:338-341.
16. Boye GI and Ampufo O (1983). Proceedings' on the first international seminar on cryptolepic (ends Boakye Yiadom k Bamgbose, S.O.A) (University of Kumasi, Ghana)
17. Karou D, Savadogo A, Canini A, Yameogo S, Montesano C, Simpore J, Colizzi V and Traore AS. Antibacterial activity of alkaloids from *S.acuta*. *African journal of biotechnology*, 2006, 5(2):195-200.
18. Garnham PCC. *Malaria parasites and other haemosporidia*. Bookwell Scientific Publications, Oxford, 1966.
19. Okwu DE and Josiah C. Evaluation of the chemical composition of two Nigerian medicinal plants Africa. *J. Biotechnology*, 2006, 5:357-361
20. Ayitey SE and Addae ML. Phytochemical nutritional and medical properties of some leafy vegetables consumed by Edo people of Nigeria. *J.Pharmacol.Drug Res*, 1977, 4:7-8.
21. Maobe MAG, Gatebe E, Gitu L and Rotich H. Preliminary phytochemical screening of eight selected medicinal herbs used for the treatment of diabetes, malaria and pneumonia in Kisii region, southwest Kenya, *European journal of applied sciences*, 2013, 5(10) : 01-06.
22. Just MJ, Recio MC, Giner RM, Cueller MU, Manez S, Billia AR and Rios JL. Antiinflammatory activity of unusual lupine saponins from *Bupleurum fruticosens*. *Thieme-E Journals*, 1998, 64:404-407.
23. Foster S and Duke JA. *Afield guide to medicinal plants*. Houghton mifflin Co, Boston, 1990.
24. Chiej R. *Encyclopedia of medicinal plants*. MacDonald publishers:London, 1984.
25. Samatha T, Shyamsundarachary R, Srinivas P, and Swamy RS. Quantification of total phenolic and total flavonoids contents in extracts of *Oroxylum Indicum* L.Kurz, *Asian Journal of Pharmaceutical and Clinical Research*, 2012, 5(4):177-179.
26. Namki M. Antioxidants and antimutagens in food. *Crit.Rev. Food Sci.Nutri*, 1990, 29:273-300.
27. Marjorie MC. Plant Products as Antimicrobial Agent. *Clin. Microbiol*, 1999, 564-582.
28. Yadav RNS and Agarwala M. Phytochemical analysis of some medicinal plants.*Journal of phytology*, 2011, 3(12):10-14.
29. Ghasemzadeh A and Ghasemzadeh N. Flavonoids and phenolic acids: Role and biochemical activity in plants and human. *Journal of medicinal plants research*, 2011 5(31):6697-6703.
30. Sharma DK. Pharmacological properties of flavonoids including flavonolignans-integration of petrocrops with drug development from plants. *Journal of scientific and industrial research*, 2006, 65:477-484.
31. Argal A and Pathak AK. CNS activity of *Calotropis gigantean* roots. *Journal of Ethnopharmacology*, 2006, 19:425-428.
32. Akiyama H, Fujii K, Yamasaki O, Oono T, Iwatsuki K. "Antibacterial action of several tannins against *Staphylococcus aureus*". *J. Antimicrobe*, 2001.

33. Bajal YPS. *Medicinal and aromatic plants*. Biotechnology in agriculture and forestry. Berlin: Springer-Verlag, 1988, Vol.24.
34. Chung KT, Wong YT, Wei CI, Huang YW and Lin Y. Tannins and human health. *Critical Reviews in Food Science and Nutrition*, 1998, 38(6): 421-464.
35. Schatzmann HJ and Rass B. Inhibition of the active Na-K-transport and Na-K-activated membrane ATPase of erythrocytes stroma by Ovabain. *Helv. Physiol. Pharmacol*, 1965, 65:47-49.
36. Newman RA, Yang P, Pawlus AD and Block KI. *Cardiac glycosides as novel cancer therapeutic agents*, 2008, 8:36-49
37. Deganhardt J. Attracting friends to feast on foes: Engineering terpene emission to make crop plant more attractive to herbivore enemies. *Curr. Opin. Biotechnol*, 2003, 14:169-176.
38. Pichersky E and Gershezon J. The formation and function of plant volatiles, perfumes for pollinator attraction and defence. *Curr. Opin. Plant Biol*, 2002, 5:237-243.
39. Viega VF Jr, Rosas, EC, Garvalho MK, Henriques MGMO and Pinto AC. Chemical composition activity of Brazilian Copaiba oils from *Copaifera reticulata* (Ducke) and *Copaifera multifuga* (Hayne). A Comparative study. *J. Ethnopharmacol*, 2007, 112: 248-254.
40. Piera FA, Souza CF, Costa JCM, Barreto MAO, Espescheit IF, Silva VO, Moreira MAS. Inhibition of *E. coli* from mastitic milk by Agrarias, 2011, 32: 1929-1934
41. Santo AO, Ueda-Nakamura T, Dias Fiho BP, Veiga VF Jr, Pinto AC, Nakamwa CV. Antimicrobial activity of Brazilian Copaiba oils obtained from different species of the *Copaifera* genus. *Mem. Inst. Oswaldo Cruz*, 2008, 103:277-281.
42. Nero LA, Moreira MAS. Antimicrobial activity of autoclaved and non-autoclaved Copaiba oil on *Listeria monocytogenes*. *Cienc. Rural*, 2010, 40:1797-1801.
43. Leandro LM, Vargas FS. Chemistry and Biological activities of terpenoids from Copaiba (*Copaifera SPP*). *Oleoresins*, 2012, 17:3866-3889
44. Lima SRM, Veiga VF Jr, Christo HB, Pinto AC, Fernandes PD. In vivo studies on the anticancer activities of the *Copaifera multifuga* (Hayne) and its fractions. *Phytother. Res*, 2003, 17:1048-1052.
45. Geris R, Silva IG, Silva HHG, Barrison AG. Diterpenoids from *Copaifera reticulata* Ducke with larvicidal activity against *Aedes aegypti* (L): Diptera (Culicidae). *Rev. Inst. Med. Trop. S. Paulo*, 2003, 50:25-28.

Source of support: Nil

Conflict of interest: None Declared