

Original Research Article

MONOPHYLY OF FOUR SECTIONS OF GENUS DENDROBIUM (ORCHIDACEAE): EVIDENCE FROM NUCLEAR RIBOSOMAL DNA INTRENAL TRANSCRIBED SPACER (ITS) SEQUENCES

Maryam Moudi^{1*} and Rusea Go^{2,3}

¹Department of Biology, Faculty of Science, University of Birjand, Birjand, South Khorasan, Iran ²Department of Biology, Faculty of Science, Universiti Putra Malaysia, 43400 UPM Serdang, Selangor Darul Ehsan, Malaysia ³Institute of Tropical Forestry & Forest Products, Universiti Putra Malaysia, 43400 UPM, Serdang, Selangor Darul Ehsan, Malaysia

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Abstract: Evolutionary relationship among four sections of the genus *Dendrobium* (*Aporum, Crumenata, Strongyle* and *Bolbidium*) were inferred from nucleotide sequence variation in nuclear ribosomal DNA of Peninsular Malaysia species. The Internal transcribed spacer (ITS) of nuclear ribosomal DNA from representative of 29 species of genus *Dendrobium* and two species of genus *Bulbophyllum* as out group were analyzed by polymerase chain reaction amplification and DNA sequencing. Data analyses have been carried out using Maximum Parsimony (MP) as a character based method by different software (PAUP^{*}4. o B 10 and Mega 5). Parsimony analyses of sequences resulted in a well-resolved phylogeny. The data suggested that Internal Transcribed Spacer (ITS) is a reliable marker for the phylogenetic study of *Dendrobium* compared to previous studies using chloroplast DNA with low resolution level among sections. The results demonstrated that the four sections formed a monophyletic group and that it is best to recognise only one section instead of four. Based on ICBN rules, the name *Aporum* has a priority to be used for this new classification.

Key Words: Dendrobium, Orchidaceae, monophyly, nuclear ribosomal DNA, Maximum Parsimony.

INTRODUCTION

With around 800-1500 species worldwide, Dendrobium Sw. is one of the three largest genera in family Orchidaceae¹. This genus was first recognized by Olofswartz in 1799. The name comes from the Greek words, Dendron meaning tree and bios for life; it means that the plant which lives in trees. Most species of Dendrobium are epiphytes in primary forest, less often lithophytes; only very few are terrestrials². Orchids are cosmopolitan in distribution, occurring in every habitat, found all over the world except in the coldest and driest regions³. Dendrobium like other orchids are generally found in wetter conditions such as the tropical regions with high annual rainfall and without too much seasonal variation throughout the year⁴. The great majority are found in the tropics, mostly in Asia, South America, and Central America. For Example, in Malaysia, the Dendrobium thrives well with this condition, a mean rainfall of about 2,500 mm and the lowest of about 1, 750 mm⁵. Dendrobiums are one of the most popular orchids due to their medicinal and commercial values³. Some species like D. bifarium Lindl., D. planibulbeLindl., & D. purpureum (Nichols) Dockr. are used in Malesia as medicinal Herbs to treat skin disorders. They have floriferous flower sprays that come in a wide variety of colors, sizes and shapes, available throughout the year and the blooms last several weeks to months⁶.

Traditionally, comparative vegetative anatomy and plant systematics were two general strategies to assess the relationships among the taxa in *Dendrobium*⁷. Problems associated with variability and

*Corresponding Author:

Maryam Moudi, Department of Biology, Faculty of Science, University of Birjand, Birjand, South Khorasan, Iran. plant growth conditions are easy to cause confusion in identification. The the species widespread development of molecular techniques for genetic analysis in the past decade has led to the increase of knowledge of orchid genetic diversity. The common molecular data used in plant systematics comes from two sources: chloroplast DNA (cpDNA) and nuclear ribosomal DNA (nrDNA)⁸. The nuclear ribosomal RNA (rRNA) genes involved 18S (small subunit), 28S (large subunit) and 5.8S, are separated by two internal transcribed spacer regions ITS1 and ITS2 (Figure 1). In eukaryotic genome, these genes are repeated in several hundred or perhaps thousands of copies⁹. The nuclear rDNA internal transcribed spacer (ITS) has become the most valuable region in plant molecular studies, especially in lower taxonomic level, since it was first used in phylogenetic inference by Baldwin in 1992¹⁰. As the nuclear gene region, it is biparentally inherited and has higher rates of base substitution. Indeed, this region can be amplified easily using standard PCR methods, because it exists in high copy number and priming sites surrounding the 18S and 26S regions are highly conserved¹¹. Although usage of ITS sequence data is very common in phylogenetic studies, the characteristics of nrDNA evolution may result in a comparison of paralegals if sufficient caution is not exercised¹². In 2003, Clements conducted а phylogenetic study on sub-tribe Dendrobiinae using ITS¹³. The genetic relationship of 36 Dendrobium species in China was determined using sequence analysis of the internal transcribed spacer (ITS) region of ribosomal DNA by Yuan and his co-workers in 2009.



Table 1: List of studied taxa

In their study, the phylogenetic relationships made by ITS DNA analysis incompletely supported previously published morphological data. Molecular evidence from these studies recommended that the genus Dendrobium is polymorphic with a complex genetic background at the species level¹⁴. In Peninsular Malaysia, Dendrobium is the second largest genus of the Orchidaceae whereby it has 16 sections and 86 species3. A total of 28 sections of Dendrobium has been recorded from Malesia by the National Herbarium of Netherland (Source: http:// www.national herbarium. nl/pubs/orchid web/genera/Dendrobium). In this study, The main objectives of the present study are to determine the phylogenetic relationship between the four sections of the genus Dendrobium (Aporum, Crumenata, Strongyle, and Bolbidium) based on nrDNA (ITS) in Peninsular Malaysia.



Figure 1: A sketch of the nuclear ribosomal RNA (rRNA) genes showing relative positions of rRNA genes subunit separated by the Internal Transcribed Spacer region (ITS).

	Species	Section	Location	Gene Bank Accession Number
1	Dendrobium aloifolium	Aporum	UPM Green house, No.5	KC507775
2	Dendrobium grande	Aporum	Labuk Tapah, Selai, PM	KC507779
3	Dendrobium leonis	Aporum	UPM Green house, No.5	KC507774
4	Dendrobium quadrilobatum	Aporum	Kuala Krai, Kelantan, PM	KC507778
5	Dendrobium rosellum	Aporum	UPM Green house, No.5	KC507777
6	Dendrobium terminale	Aporum	Sungai Bertedung, Endau Rompin, PM	KC507776
7	Dendrobium clavator	Crumenata	Sungai Bertedung, Endau Rompin, PM	KC507762
8	Dendrobium crumenatum	Crumenata	Genting Highlands, PM	KC507780
9	Dendrobium setifolium	Crumenata	Sungai Bertedung, Endau Rompin, PM	KC507763
10	Dendrobium truncatum	Crumenata	Cameron Highlands, PM	KC507761
11	Dendrobium kentrophyllum	Strongyle	Fraser's Hill, PM	KC507764
12	Dendrobium singaporense	Strongyle	Cameron Highlands, PM	KC507765
13	Dendrobium subulatum	Strongyle	Gunung Nuang, PM	KC507766
14	Dendrobium pachyphyllum	Bolbidium	Fraser's Hill, PM	KC507769
15	Dendrobium hymenanthum	Bolbidium	Cameron Highlands, PM	KC507768
16	Dendrobium striatellum	Bolbidium	Kuala Krai, Kelantan, PM	KC507767
17	Dendrobium sp2	?	G. Kelapak buruk, PM	KC507770
18	Dendrobium sp3	?	UPM Green house, No.5	KC507771
19	Dendrobium sp4	?	UPM Green house, No.5	KC522821
20	Dendrobium sp5	?	UPM Green house, No.5	KC522822
21	Dendrobium sp6	?	UPM Green house, No.5	KC522823
22	Dendrobium heterocarpum	Dendrobium	NCBI	JF713105.1
23	Dendrobium nobile	Dendrobium	NCBI	JF713365.1
24	Dendrobium crepidatum	Dendrobium	NCBI	HM054624.1
25	Dendrobium fimbriatum	Dendrobium	NCBI	HM054636.1
26	Dendrobium gibsonii	Dendrobium	NCBI	HQ114256.1
27	Dendrobium farmeri	Callista	NCBI	HM054629.1
28	Dendrobium thyrsiflorum	Callista	NCBI	HM054759.1
29	Dendrobium macrophyllum	Lautoria	NCBI	AY239979.1
30	Bulbophyllum inunctum	Sestochilus	Cameron Highlands, PM	KC507773
31	Bulbophyllum macranthum	Sestochilus	Gunung Jerai, PM	KC507772

MATERIALS AND METHODS

Plant materials

Dendrobium samples (21 fresh samples) of the four sections (Aporum, Crumenata, Strongyle, and Bolbidium) were collected along the trials of the selected areas, UPM greenhouse and some nursery in Peninsular Malaysia (Table 1). There were five Dendrobium species that could not be identified because of the lack of flowers (however, their morphological characters were nearly similar to three sections Aporum, Crumenata and Strongyle. The fresh leaves or stems of species were used for DNA extraction. For the better comparison in the data analysis step, the sequences of the eight species of genus Dendrobium that belong to the three sections Dendrobium, Callista and Latouria retrieved from the NCBI database were used. In addition, two species of the genus Bulbophyllum (B. macranthum and B. inunctum) were used as outgroup.

Genomic DNA extraction, amplification, and sequencing

Genomic DNA was extracted from leaf samples using a cetyltrimethyl ammonium bromide (CTAB) method as described previously by Wang et al 2004 with minor modification¹⁵. The concentration of genomic DNA samples was determined by Nano drop machine and necessary dilutions were done, followed by verification with 0.8% agarose gel electrophoresis. The PCR conditions were 95°C 5 min; 35 cycles: 95°C 1 min; 50/55°C 2 min; 72°C 2 min; 72°C 7 min. The PCR products were separated by1% agarose gel electrophoresis and purified by Wizard SV Gel and PCR Clean-Up System (Promega, USA). After that, all of purified PCR products were sent to the First Base Company for sequencing. Finally the sequences were submitted to the NCBI database (Table 1).

DNA sequence data analysis

DNA sequences obtained from ITS region were aligned by BioEdit ver.7.0.2.¹⁶. The Maximum Parsimony (MP) method was selected for the construction of phylogenetic trees. The output data were processed using PAUP^{*} 4.0B10¹⁷ and MEGA5¹⁸ to draw the phylogenetic trees. A total of 1000 bootstrap replicates was calculated for all methods.

RESULTS AND DISCUSSION

Complete ITS sequences were analysed for 29 Dendrobium species plus Bulbophyllum inunctum and Bulbophyllum macranthum as outgroup. The aligned sequences consisted of 490 nucleotide characters. 170 characters were conserved among all taxa, 291 were variable, and 180 were parsimony informative. The analysis methods, Maximum Parsimony (MP) were used to infer the evolutionary history. Unweighted parsimony analysis resulted in 12 most parsimonious trees [tree length = 589, consistency index (CI) = 0.58, retention index (RI) = 0.71 and homoplasy index (HI) = 0.33]; One of the most parsimonious trees was shown in Figure.2.

One of the loci that is commonly used for phylogenetic inference is ITS with a high copy number and relative range of phylogenetic utility¹⁹. The use of nrDNA regions has become popular and is used in different level of taxa, especially the low level taxa, such as infrageneric species²⁰. The utility of this region in the molecular study of family Orchidaceae is very widespread. It is often used in phylogenetic studies at various taxonomic levels in most groups of the family Orchidaceae²¹; Salazar et al., 2003²²; Van den Berg et al., 2005²³; whitten *et al.*, 2007²⁴). Andre Schuiteman (2011) has used ITS to do phylogeny of genus Dendrobium. Actually, the obtained results from ITS marker in this study confirmed and extended his hypothesis, which indicated that the four sections (Aporum, Crumenata, Strongyle and Bolbidium) can be considered as one

section instead of four. The original studies by Yukawa and co-workers (1993^{25} , 1996^{26} , 2000^{27} , 2001^{28}) using DNA markers (*ma*tK and ITS) have made available a number of insights concerning the phylogeny of genus *Dendrobium* and subtribe Dendrobiinae, which were confirmed and improved by later studies (Clements 2003^{13} , 2006^{29}).



Figure 2: One of most parsimonious trees of selected *Dendrobium* sections based on nrITS sequence data. A (1-2) and B are mainclades and include the different sections of genus *Dendrobium*. A1 (I and II) denote the four sections *Aporum*, *Crumenata*, *Strongyle* and *Bolbidium* form a monophyletic group and can be considered as one section named *Aporum*.

In this study, several attempts were made to amplify ITS from a number of Dendrobium species. The lower level of homoplasy and around 37% PIC was established in this region to show a high level of branch support in data analysis. Furthermore, it confirmed that sections Crumenata, Bolbodium, Aporum and Strongyle formed a well-supported monophyletic group. That it is best to recognise only one section instead of four. Based on ICBN rules, the name Aporum has a priority to be used for this new classification. It also suggested that each three sections Aporum, Crumenata and Strongyle were not monophyletic, whereas section Bolbidium was monophyletic with strong support is around 99%. Furthermore, the results showed the unidentified Dendrobium excluding D. sp6 were nested with species of two sections Aporum and Strongyle; therefore they may be belonged to these two sections.

The species *D.* sp2 was close to *D. singaporense* (*Strongyle*) with bootstrap percentage of more than 50%. The two species *D.* sp4 and *D.* sp5 were close together with strong support (BP of more than 90%). Also, based on the results *D.* sp3 was related to the section *Aporum*. This species was close to *Dendrobium grande* but with not strong bootstrap. Although *D.* sp6 was similar to section Crumenata species, according to the morphological characters, but the result showed that this specie was close to the section *Dendrobium based* on Molecular evidence.

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REFERENCES

- Xiang XG, Schuiteman A, Li DZ, Huang WC, Molecular systematic of Dendrobium (Orchidaceae, Dendrobieae) from mainland Asia based on plastid and nuclear sequences, Molecular Pylogenetics and Evolution, 2013, 69, 950-960.
- 2. Schuiteman A, Dendrobium (Orchidaceae): to split or not split?, Gardens Bulletin Singapore, 2011, 1and 2, 245-257.
- 3. Seidenfaden G and Wood JJ, The Orchids of Peninsular Malaysia and Singapore. Fredensborg: Olsen and Olsen, 1992.
- 4. Wanga HZ, Fenga SG, Lua JJ, Shia NN, Liub JJ, Phylogenetic study and molecular identification of 31 Dendrobium species using inter-simple sequence repeat (ISSR) markers, Scientia Horticulture, 2009, 122, 440-447.
- 5. Segerbäck LB, Orchids of Malaya, Malaysia: Balkema, 1992.
- 6. Lavarack PS, Harris W and Stocker G, Dendrobium and its relatives. Oregon, Portland: Timber Press, 2000.
- 7. Adams PB, Burke JM and Lawsen SD. Systematic analysis of Dendrobium Swartz section Dendrocoryne in the Australian region, Plant systematic Evolution, 2006, 260, 65-80.
- 8. Small RL, Cronn RC and Wendel JF, LAS. JOHNSON REVIEW No. 2. Use of nuclear genes for phylogeny reconstruction in plants, Australian systematic Botany, 2004, 17, 145-170.

- 9. Hills DM, Moritz C, Mable, BK, Molecular systematics, Sunderland, MA: Sinauer Associate, Inc, 1996.
- 10. Baldwin BG, Phylogenetic utility of the internal transcribed spacers of nuclear ribosomal DNA in plants: an example from Compositae, Molecular Phylogenetics and Evolution, 1992, 1, 3-16.
- White TJ, Bruns T, Lee S, Taylor J, Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics (M. A. Innis, D. H. Gelfand, J. J. Sninsky, and T. J. White Ed.). San Diego, California, USA: Academic Press, 1990.
- 12. Alvarez I, Wendel JF, Ribosomal ITS sequences and plant phylogenetic inference, Molecular Phylogenetics and Evolution, 2003, 29 (3), 417–434.
- 13. Clements MA, Molecular phylogenetic systematics in the Dendrobiinae (Orchidaceae), with emphasis on Dendrobium section Pedilonum, Telopea, 2003, 10 (1), 247-272.
- 14. Yuan, ZQ, Zhang JY and Liu T, Phylogenetic relationship of China Dendrobium species based on the sequence of the internal transcribed spacer of ribosomal DNA, Biological Plantarum, 2009, 53 (1), 155-158.
- 15. Wang HZ, Wang YD, Zhou XY, Ying QC, Zheng KL, Analysis of genetic diversity of 14 species of Cymbidium based on RAPDs and AFLPs, Acta Biologiae Exprimetalis Sin, 2004, 37 (6), 482–486.
- Hall AT, Bio Edit: a user friendly biological sequence alignment editor and analysis performer windows 95/98/NT, Nucleic Acid Symposium series, Axford Journals, 1999, 41, 95-98.
- 17. Swofford DL, PAUP. Phylogenetic analysis using parsimony, Version 4b10. Sinauer, Sunderland, Massachusetts, USA,2002.
- Tamura, K, Peterson D, Peterson N, MEGA5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods, Molecular Biology and Evolution, Advance access: 10.1093/molbev/msr121, 2011.
- 19. Soltis DE, and Soltis PS, Molecular systematics of plants II: DNA sequencing,Boston: Kluwer Academic Publishers, 1998.

- Choi HK, Kim CK, Shin HC, Molecular reexamination of Korean Umbelliferae based on internal transcribed spacer sequences of rDNA: Ligusticum tenuissimum (Nakai) Kitagawa, and Libanotis coreana (Wolf)vKitagawa. Plant Biology,2000, 43, 136-142.
- 21. Koehler K, Stechele M, Hetzel U, Domingo M, Schönian G, Zahner H, *et al.*, Cutaneous leishaniosis in a horse in Southern Germany caused by Leishmania infantum, Veterinary Parasitology, 2002, 109 (1-2), 9-17.
- 22. Salazar GA, Chase MW, Soto Arenas MA, Ingrouille M, Phylogenetics of Cranichideae with emphasis on Spiranthinae (Orchidaceae: Orchidoideae): Evidence from plastid and nuclear DNA sequences, American Journal of Botany, 2003, 90 (5), 777-795.
- 23. Van Den Berg C, Goldman DH, Frudenstein JV, Pridgeon AM, Cameron KM and Chase MW, An overview of the phylogenetic relationships within Epidendroideae inferred from multiple DNA regions and recircumscription of Epidendreae and Arethuseae (Orchidaceae), American Journal of Botany, 2005, 92(4), 613-624.
- 24. Whitten WM, Blanco MA, Williams NH, Koehler S, Carnevali G, Singer RB. Molecular phylogenetics of Maxillaria and related genera (Orchidaceae: Cymbidieae) based on combined molecular datasets, American Journal of Botany, 2007, 94(11), 1860-1889.

- 25. Yukawa T, Kurita S, Nishida M, Hasebe M, Phylogenetic implications of chloroplast DNA restriction site variation in subtribe Dendrobiinae (Orchidacaeae), 1993, Lindleyana, 8, 211–221.
- 26. Yukawa T, Ohba H, Cameron KM, Chase MW, Chloroplast DNA phylogeny of subtribe Dendrobiinae (Orchidaceae): insights from a combined analysis based on rbcL sequences and restriction site variation. Journal Plant Research, 1996, 109 (2), 169–176.
- 27. Yukawa T, Kita K, Handa, T, DNA phylogeny and morphological diversification of Australian Dendrobium (Orchidaceae), In: Wilson, K.L. and Mossison, D.A. (eds) Monocots: Systematics and Evolution, Melbourne: CSIRO Publishing, 2000,465–471.
- 28. Yukawa T, Molecular phylogeny of Dendrobium, In: Nagata, H. and Ichihashi, S. (Eds) Proceedings of the seventh Asia Pacific orchid conference (APOC7), 2001.
- 29. Clements MA, Molecular phylogenetic systematics in Dendrobieae (Orchidaceae), Aliso, 2006, 22: 465–480.

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