

ISOTHIOCYANATES: NATURALLY OCCURRING HepG2 HUMAN LIVER CARCINOMA INHIBITOR

Bansal P¹, Medhe S¹, Ganesh N² and Srivastava MM^{1*}

¹Department of Chemistry, Dayalbagh Educational Institute, Dayalbagh, Agra-282110, India ²Department of Research, Jawaharlal Nehru Cancer Hospital & Research Centre, Bhopal-462001, India

***Corresponding Author:** Dr. Srivastava MM, Department of Chemistry, Dayalbagh Educational Institute, Dayalbagh, Agra-282110, India

Received for publication: October 19, 2012; Accepted: December 02, 2012.

Abstract: In vitro anticancer screening of different isothiocyanates (ITCs) [Allyl isothiocyanate (AITC), Phenylethyl isothiocyanate (PEITC) and Sulphoraphane (SUL)], derived from naturally occurring *Eruca* sativa seed oil (SO) against HepG2 human liver carcinoma cell line has been carried out using MTT, TBE and SRB bioassays. Combination of ITCs, in ratio (1:1:1) exhibited maximum percentage inhibition of 97.12%, 96.34% and 96.56% by MTT, TBE and SRB respectively against reference drug doxorubicin at concentration 50 μ M. Percent inhibition has been found to be more pronounced in isothiocyanates combination compared to its natural source *E. sativa* seed oil.

Keywords: Carcinoma, Eruca sativa, Isothiocyanates

INTRODUCTION

Antioxidants protect the body from destructive free radicals, which are generated when the body uses oxygen to make energy and from exposure to environmental factors. Left unchecked, free radicals cause uncontrollable chemical reactions that damage the body. Without the help of antioxidants, decay would soon destroy human bodies by damaging DNA and accelerating aging. Experimental findings indicate that antioxidants inhibit the growth of cancer cells or directly kill them (1). Research also indicates that antioxidants may work with chemotherapy to kill more cancerous cells than approach alone (2). Since it is known that antioxidants work together (3), a combination of antioxidants can be expected to have an even greater anticancer effect (4). A combination of antioxidants has been found to be more effective in reducing the growth of cancer cells than the individual antioxidant (5; 6). One major benefit of antioxidants is the fact that they do not interfere with the way that chemotherapy produces its anticancer effect. The use of antioxidants with chemotherapy showing no interference has been reported (7).

E. sativa, (Miller) is the most representative species of annual herb, which originated in the Mediterranean region (8), now is widely distributed all over the world, particularly in India (9), where the seeds are used for the production of a traditional spicy (taramira) oil. *E. sativa* oil is brought to the attention of the scientific community for various pharmacological properties viz. antidiabetic, antirenal-failure, antigastro-intestinal indigestion and antineurotoxic activity (10).

In continuation of our work (11; 12; 13; 14), on screening of Indian medicinal plants for various

pharmacological activities, we have recently shown that *E. sativa* is a relevant source of antimicrobial activity (15). Various isothiocyanates have been isolated from *E. sativa* seed oil (16). The combinational effects of the isothiocyanates derived from *E. sativa* seed oil have been studied on B16F10 mice melanoma cell line (17). Encouraging results on the above lines have inspired us to explore the *invitro* effect of combination of three isothiocyanates (Allyl isothiocyanate, Phenylethyl isothiocyanate and Sulphoraphane) for anticancer activity against HepG2 human liver carcinoma cell line and to compare its efficacy with *E. sativa* seed oil.

MATERIALS AND METHOD

Chemicals and Reagents:

Dulbecco's modified eagle's medium (DMEM), fetal bovine serum (FBS), glutamine, penicillin and streptomycin were purchased from Gibco BRL, USA. Trypan blue dye, MTT [3-(4, 5- dimethyl thiazol-2-yl)-2, 5diphenyl tetrazolium] dye and doxorubicin (Trade name: Adriamycin) were obtained from Sigma-Aldrich Chemical Company (UK). Allyl isothiocyanate, phenylethyl isothiocyanate and Sulphoraphane were used as test samples. Analytical standards were purchased from Fluka-Sigma-Aldrich. Double distilled water was used throughout the experiment. All other chemicals and solvents used were of highest purity commercially available (Sigma–Fluka–Aldrich) and were of analytical grade.

Cell culture:

HepG2 human liver carcinoma cells were purchased from National Centre for Cell Sciences, Pune, India. These cells were maintained in DMEM



medium supplemented with antibiotics, L-glutamine (2 mM) and 10% fetal calf serum. HepG2 human liver carcinoma cells were considered for in vitro cytotoxic studies on the basis of their metastatic nature. The cells were grown in the following conditions: CO_2 (5%) atmosphere in high humidity (95%) at 37°C in a CO_2 incubator. Each batch of cells was assessed for cell cytotoxicity by TBE, SRB and MTT assays.

Trypan Blue Exclusion Assay:

Cells (1×10⁵/plate) were incubated in poly-l-lysine precoated tissue culture petri plates with complete medium (MEM medium with fetal bovine serum) and allowed to adhere for 24 h in CO_2 incubator at 37 °C. The medium was replaced with incomplete medium (MEM medium without fetal bovine serum) containing serial dilution series of test samples $(3 - 50 \mu M)$ separately for 24 h in CO₂ incubator at 37 °C. The medium was removed from the wells by aspiration and trypsinized to obtain the cell suspension. Trypan blue dye (0.4%; 0.1 ml) was mixed with cell suspension, 15 min prior to completion of incubation period (18). At the end of incubation period, the number of viable cells (unstained) was counted using a haemocytometer. Viability was expressed as a percentage of control number of cells excluding trypan blue dye.

Methyl Thiazole Tetrazolium Assay:

MTT assay depends on the mitochondrial enzyme reduction of tetrazolium dye to determine cell viability. Briefly, the cells were plated at a density of 1×10⁵ cells/well into 96- well plates and allowed to adhere for 24 h in CO_2 incubator at 37 °C. The medium was replaced with the serum free medium containing different concentrations of test samples $(3 - 50 \mu M)$ again for 24 h in CO₂ incubator at 37°C. MTT dye (20μ l; 5 mg/ml) was added to each well 4 h prior to completion of incubation period. The medium was removed by adding 200 μ l of DMSO to each well and incubated for further 10 min (19). The absorbance was read at 550 nm using ELISA reader (Synergy HT, Biotech, USA). The average values were determined from triplicate. Percent inhibition was calculated by using the formula: (C-T)/C×100, where C=Absorbance of control, T=Absorbance of Treatment. The IC₅₀ values of test compound were compared with standard drug (20).

SRB Assay:

Cells ($1x10^{5}$ /well) were seeded in poly-l-lysine precoated flat bottomed 96 well tissue culture plates and allowed to adhere for 48 h in CO₂ incubator at 37°C. The medium was replaced with the complete medium containing different concentrations of test samples (3 -50 µM) separately again for 24 h in CO₂ incubator at 37°C. The plates were stained with sulphorhodamine B solution (0.4% in water) in dark for 30 min and then excess dye was washed out with acetic acid (1%) (21).The unwashed dye was eluted with tris-buffer and quantified at 570 nm using a microplate reader (Synergy HT, Biotech, USA).

Statistical Analysis:

Data are expressed in percent inhibition with respect to the control. The percentages of cell inhibition were used to determine the IC_{50} values. All experimental data were given as mean ± SD. Statistical analysis was carried out using the one-way analysis of variances (ANOVA). Post Dunnett test was applied between control, reference drug and test samples using Graph Pad Prism software. Probability values were found to be less than 0.05 (p < 0.05).

RESULTS AND DISCUSSION

A combinational study of three different Isothiocyanates: Allyl isothiocyanate, Phenylethyl isothiocyanate and Sulphoraphane were screened for their *in vitro* cytotoxic activity against HepG2 human liver carcinoma cell line. Inhibitory concentration (IC_{50}) was evaluated for all the three native phytoproducts with respect to vehicle control and standard drug (doxorubicin). Based on (IC_{50}) values, different combinations of the three phytoproducts were prepared and monitored for their cytotoxic effect against HepG2 human liver carcinoma cell line (Fig 1).



Each value is mean ± SD (n=3). P<0.01 vs control. Control: 2% dimethyl sulphoxide, AITC: Allyl isothiocyanate, PEITC: Phenylethyl isothiocyanate, SUL: Sulphoraphane



Fig. 1: *In vitro* cytotoxic effect of Isothiocyanates (single and combination) on HepG2 human liver carcinoma cell line using

(a) MTT

(b) TBE and

(c) SRB bioassays.

Cytotoxic activity of different Isothiocyanates and E. sativa seed oil were screened against HepG2 human liver carcinoma cell line with increasing concentrations after 48 h incubation first by the MTT, TBE followed by SRB bioassay (22). Current assays for measuring cytotoxicity are based on alterations of plasma membrane permeability and the consequent release of components into the supernatant or the uptake of dyes. In case of trypan blue exclusion assay, the dead cells uptake the dye while the viable cells are excluded (23). In MTT assay, dead cells are unable to metabolize yellow tetrazolium salt while viable cells metabolize yellow tetrazolium salt into purple formazan crystal (24) and sulphorhodamine B is an anionic aminoxanthene dye that forms an electrostatic complex with the basic amino acid residues of proteins under moderately acidic conditions, which provides a sensitive linear response (25). Three (MTT, TBE and SRB) bioassays are likely to provide data of percent inhibition of phytoproducts and hence selected for the present study.

Table.1 represents the data of percent inhibition of Doxorubicin (reference drug). AITC+PEITC+SUL (isothiocyanates combination) and SO (2mg/ml) against HepG2 human liver carcinoma cell line by the MTT, TBE and SRB bioassays. A perusal of the data highlights maximum percent inhibition of AITC-77.25%, PEITC-65.38%, SUL-63.85%, AITC+PEITC-86.19%, PEITC+SUL-85.6%, SUL+AITC-84.39%, AITC+PEITC+ SUL-97.12, SO (2mg/ml)- 68.46% for MTT assay; AITC-77.39%, PEITC-64.28%, SUL-63.49%, AITC+PEITC-84.75%, PEITC+SUL-84.39%, SUL+AITC-83.69%, AITC+PEITC+SUL-96.34, SO (2mg/ml)- 66.82% for TBE assay; AITC-77.85%, PEITC-63.42%, SUL-62.35%, AITC+PEITC-84.68%, PEITC+SUL-84.39%, SUL+AITC-83.57%, AITC+PEITC+SUL-96.56, SO (2mg/ml)- 65.92% for SRB assay at concentration of 50 µM. Percent inhibition resulting from TBE, SRB and MTT bioassays demonstrated that seed oil and

isothiocyanates exhibit significant cytotoxicity against HepG2 human liver carcinoma cell line. However, combination of isothiocyanates showed more pronounced efficacy (maximum inhibition 97.12%) at the concentration 50 μ M at 24 h compared to *E sativa* seed oil against HepG2 human liver carcinoma cell line (p < 0.05).

Table 1: In vitro cytotoxic effect of	of Doxorubicin,	ITCs		
combination and Seed oil on H	HepG2 human	liver		
carcinoma cell line using MTT, TBE & SRB bioassays				

Samples	Bioassays (% inhibition)			
(µM)	МТТ	TBE	SRB	
Control	01.62±0.83	01.85±0.95	01.22±0.33	
DOXO				
3	54.25±1.56	53.86±1.45	54.86±1.36	
10	71.21±1.61	71.68±1.52	71.21±1.28	
30	97.54±1.65	96.65±1.63	96.54±1.38	
AITC+PEITC+SUL				
3	22.65±1.58	25.39±1.47	27.39±1.49	
10	43.17±1.51	41.25±1.49	42.59±1.52	
30	63.75±1.54	62.87±1.35	63.32±1.56	
50	97.12±1.64	96.34±1.46	96.56±1.55	
SO (2mg/ml)				
3	9.53±1.62	9.51±1.52	9.34±1.54	
10	23.43±1.58	22.59±1.53	22.68±1.56	
30	49.59±1.54	48.37±1.56	47.16±1.59	
50	68.46±1.64	66.82±1.46	65.92±1.51	

Each value is mean \pm SD (n=3). p<0.01 vs control. Control: 2% dimethyl sulfoxide, Doxo: Doxorubicin, SO: Seed oil, AITC: Allyl isothiocyanate, PEITC: Phenylethyl isothiocyanate, SUL: Sulphoraphane.

CONCLUSION

The present piece of work demonstrates that the combination of three isothiocyanates isolated from *E. sativa* seed oil shows more significant in vitro cytotoxic effect compared to its naturally occurring seed oil. The detailed *in vivo* studies are in progress. It is recommended to understand specific cellular, molecular and genetic mechanisms of combinational effects of these phytoproducts that contribute cancer growth and progression.

ACKNOWLEDGEMENTS

The authors gratefully acknowledge Prof. V.G Das, Director, Dayalbagh Educational Institute, Dayalbagh, Agra for providing necessary facilities. Ms PB acknowledges University Grants Commission, New Delhi- Basic Scientific Research (UGC-BSR) for providing research fellowship.

REFERENCES

1. Salganik R, The benefits and hazards of antioxidants: controlling apoptosis and other protective mechanisms in cancer patients

and the human population. Journal of the American College of Nutrition. 2001, 20(5), 464S-472S.

- 2. Maiti AK, Elevate the ROS level to kill cancer cells during chemotherapy. Chemotherapy. doi:10.4172/2167-7700.1000e119, 2012, 1 (5)
- Medhe S, Bansal P & Srivastava MM, Enhanced antioxidant activity of gold nanoparticle embeded 3, 6 dihydroxyflavone: A combinational study. Applied Nanosciences. doi 10.1007/s 13204-012-0182-9, 2012
- 4. Sagar SM, Should patients take or avoid antioxidant supplements during anticancer therapy? An evidence-based review. Current Oncology. 2005, 12(2), 44-54
- 5. Prasad KN, Kumar A, Kochupillia V & Cole WC, High doses of multiple antioxidant vitamins: essential ingredients in improving the efficacy of standard cancer therapy. Journal of the American College of Nutrition. 1999, 18(1), 13-25
- 6. Prasad KN & Kumar R, Effect of individual and multiple antioxidant vitamins on growth and morphology of human non tumorigenic and tumorigenic parotid acinar cells in culture. Nutrition and Cancer. 1996, 26,11-19
- Block KI, Koch AC, Mead MN, Tothy PK, Newman RA & Gyllenhaal C, Impact of antioxidant supplementation on chemotherapeutic efficacy: A systematic review of the evidence from randomized controlled trials. Cancer Treatment Reviews. doi:10.1016/j.ctrv.2007.01.005, 2007
- 8. Steven JT, Jackson & Singletary KW, Sulforaphane: a naturally occurring mammary carcinoma mitotic inhibitor which disrupts tubulin polymerization. Carcinogenesis, 2004, 25, 219-227
- 9. Park EJ & Pezzuto JM, Botanicals in cancer chemoprevention. Cancer and Metastasis Reviews. 2002, 21,231–255
- Kataya HAH & Hamza AA, Red cabbage (Brassica oleracea) ameliorates diabetic nephropathy in rats. Evidence-Based Complementary and Alternative Medicine. 2008, 5(3),281-287
- Ojeswi BK, Khoobchandani M, Hazra DK & Srivastava MM, Invitro antibreast cancer efficacy of two indigenous plants on human cancer cell line MCF-7. National Academy Science Letters. 2009, 32,105-109
- Khoobchandani M, Ojeswi BK, Sharma B & Srivastava MM, Chenopodium album preventive progression of cell growth and enhancement in cell toxicity in human breast cancer cell lines. Oxidative Medicine and cellular longevity. 2009, 2(3), 160-165
- Ojeswi BK, Khoobchandani M, Hazra DK & Srivastava MM, Protective effect of Thuja occidentalis against DMBA-induced breast cancer with reference to oxidative stress. Human and Experimental Toxicology. 2010, 29(5), 369-375

- Khoobchandani M, Bansal P, Medhe S, Ganesh N & Srivastava MM, Antioxidant and antimutagenic activities of isothiocyanates rich seed oil of Eruca sativa plant chemistry of phytopotentials: Health, Energy and Environmental Perspectives. DOI : 10.1007/978-3-642-23394-4_10, Springer-Verlag, Berlin, Heidelberg, 2012, 47-51
- 15. Khoobchandani M, Ojeswi BK, Ganesh N, Srivastava MM, Gabbanini S, Matera R, Iori R & Valgimigli L, Antimicrobial properties and analytical profile of traditional Eruca sativa seed oil: Comparison with various aerial and root plant extracts. Food Chemistry. 2010, 120, 217-224
- Khoobchandani M, Ganesh N, Gabbanini S, Valgimigli L & Srivastava MM, Phytochemical potential of Eruca sativa for inhibition of melanoma tumor growth. Fitoterapia. 2011, 82, 647-653
- 17. Bansal P, Medhe S, Ganesh N & Srivastava MM, Eruca sativa seed oil as promising antioxidant and anticarcinogenic dietary bioagent: Role of Isothiocyanates. National Academy Science Letters (communicated). 2012
- Frieauff W, Hartman A & Suter W, Automatic analysis of slides processed in the Comet assay. Mutagenesis. 2001, 16, 133-137
- Lee SE, Ju EM & Kim JH, Antioxidant activity of extracts from Euryale ferox seed. Experimental and Molecular Medicine. 2002, 34(2), 100–106
- Kumar V & Yadav SK, Plant-mediated synthesis of silver and gold nanoparticles and their applications. Journal of Chemical Technology & Biotechnology. 2009, 84, 151-157
- 21. Choi EH, Chang HJ, Cho JY & Chun HS, Cytoprotective effect of anthocyanins against doxorubicin induced toxicity in H9c2 cardiomyocytes in relation to their antioxidant activities. Food and Chemical Toxicology. 2007, 45, 1873-1881
- 22. Medhe S, Bansal P, Roy SK, Rajan MGR & Srivastava MM, Nanotech and Combinational Therapeutic Exploitation of Phytochemicals: Promising Chemopreventing Agent. Bionanoscience (In press), 2012
- 23. Baliga R, Zhang Z & Shah SV, Role of cytochrome P-450 in hydrogen peroxide-induce cytotoxicity to LLC-PK1 cells. Kidney International. 1996, 50, 11-18
- 24. Kumar V, Yadav SK, Plant-mediated synthesis of silver and gold nanoparticles and their applications. Journal of Chemical Technology & Biotechnology. 2009, 84, 151-157
- 25. Perez RP, CytoScan-SRB cell cytotoxicity assay. European Journal of Cancer. 1993, 29, 395-399

Source of support: Nil Conflict of interest: None Declared