



Effect of salinization on health benefits of fenugreek seedlings

Bardees Mohammad Mickky*, Muhammad Ali Abbas and Nada Mohamed Sameh Botany Department, Faculty of Science, Mansoura University, Mansoura, Egypt

Received: 14th January 2017; Accepted: 18th January 2017; Available online: 1st February 2017

Abstract: The current investigation was conducted to evaluate the potency of salinity to enhance the nutraceutical properties of fenugreek (*Trigonella foenum-graecum* L.) seedlings. Gradual doses of NaCl solution (0, 50, 100, 150 and 200 mM) were applied and the nutritional value of seedlings was determined 2 and 5 days' post-germination. Moreover, the antioxidant and antimicrobial activity along with the phenolic content of the considered seedlings were assessed. The obtained results manifested that salt application to different levels could increase the amount of protein, N, Zn and Cu in the 2- and 5- day old seedlings. Salinity could also raise the amount of vitamin C, B1, B2 and B9 but only in one stage. Conversely, the amount of reducing, non- reducing and total sugars as well as that of Fe declined under stress. Compared with their unsalted synonyms, water and methanolic extracts of salt- treated fenugreek seedlings had higher H₂O₂- scavenging activity in the two stages, while their reducing power and free radicles- scavenging activities were promoted in the second stage only. Regarding the antimicrobial activity, water and methanolic extracts of the stressed seedlings generally exhibited better antimicrobial activity against some pathogenic microbes especially *Erwinia carotovora, Bacillus subtilis* and *Candida albicans*. Only in the second stage, most of the employed salt concentrations increased the titer of tannins, saponins, alkaloids, flavonoids and total phenols in the aqueous and alcoholic extracts. Therefore, it is recommended to salinize young fenugreek seedlings to improve its health benefits in a simple, low-cost and low- risk approach.

Key words: fenugreek seedlings; salinity; nutritional value; antioxidant activity; antimicrobial potency; phenolics

Introduction

Fenugreek (Trigonella foenum-graecum L.) is an annual herb belonging to family Fabaceae. As many as 260 species of genus Trigonella were identified with the species foenum-graecum most distributed throughout the world. It is cultivated as a semi- arid crop in Mediterranean countries, Europe and Asia [1]. According to Lust [2], fenugreek is one of the oldest known medicinal plants in the recorded history. Fenugreek leaves and seeds have been famed for their anti- diabetic, cholesterollowering, antitumor, antibacterial, antifungal, vermicidal and antioxidant activities [3]. Fenugreek seeds are also reputed for numerous medicinal virtues with tonic, demulcent, emollient, carminative, emmenagogue, diuretic, aphrodisiac and expectorant properties. Moreover, they have been widely used since millennia as a treatment for stomach irritation, mouth ulcers and chapped lips [4].

With the dramatic increase in population number all over the world along with the spread of diseases, there is an ever-increasing need to cultivate medicinal plants. Yet, different forms of environmental stresses could retard agricultural activities with eventual interference with the quantitative and qualitative productivity of medicinal plants [5]. Among such stresses, salinity is one of the most problematic issues threating the agricultural sector [6]. Studies on the negative effects of salt stress on growth, development and yield of various medicinal plants could be reported [7, 8]. However, and up to our knowledge, so little attention is paid to the utilization of environmental traumas in enhancing the medicinal value of certain plants.

In recent studies, Mickky *et al.*, [9, 10] could maximize the medicinal value of alfalfa plants by growing them with decreasing water supply either alone or combined with increasing sand amount in the cultivation soil. Such stressful conditions could significantly enforce the studied plants to over- produce some medically- active phytochemicals such as phenolics, vitamins, nonphotosynthetic pigments and minerals. Furthermore, the extracts of alfalfa plants under such treatments possessed better antioxidant as well as antibacterial and antifungal activities.

Therefore, the present investigation was intended to assess the effect of gradual levels of salt stress on health benefits of fenugreek seedlings. The amount of some nutritive phytoconstituents was determined in addition to evaluating the antioxidant and antimicrobial potentialities of the extracts prepared from stressed and unstressed seedlings.

*Corresponding Author:

Dr. Bardees Mohammad Mickky
Lecturer of Plant Physiology,
Botany Department, Faculty of Science,
Mansoura University, Mansoura, Egypt.
E-mail: bardees_mickkv@mans.edu.eg



Materials and Methods

Experimental Design

Seeds of fenugreek (*Trigonella foenum-graecum* L. variety Giza 3) were obtained from Sakha Agricultural Research Center, Kafr El-Sheikh Government, Egypt. Homogeneous lot of seeds was externally sterilized using 4% NaOCI. The seeds were hydro- primed for 8 hours then allowed to germinate on gauze in darkpainted plastic boxes at 25± 2°C. The germinated seeds were thereafter sprayed with NaCl solutions at 0, 50, 100, 150 or 200 mM when required till the sampling was performed after 2 and 5 days from the experiment beginning.

Evaluation of Nutritional Value of Fenugreek Seedlings

To assess the nutritional value of fenugreek seedlings under the studied conditions; moisture, protein, nitrogen and sugar contents were determined. Also, some vitamins and heavy metals were quantified.

Determination of chemical composition:

Moisture content of fenugreek seedlings was determined according to the method described by AOAC [11]. Also, crude protein amount in the studied samples was recorded by the official Kjeldahl method of AOAC [11]. The percentage of total nitrogen could be determined then protein content was calculated using 6.25 as a coversion factor. Total sugars content was determined according to phenol- sulfuric acid method cited from Masuko *et al.*, [12]. The amount of reducing sugars was determined by dinitrosalicylic acid method of Miller [13]. The titre of non- reducing sugars could be thus calculated by subtracting the value of reducing sugars from that of total sugars.

Determination of heavy metals:

To extract heavy metals, dry plant ashes were digested in concentrated nitric acid coupled with sulfuric acid in presence of perchloric acid. After being filtered, clear plant extracts were analyzed for Fe, Zn and Cu using atomic absorption spectrophotometer (Buck Scientific Series model 214; USA) as described by Allen *et al.*, [14].

Determination of vitamins:

For determination of vitamin C, the titration routine of Ogunlesi *et al.*, [15] was followed using 2, 6-dichlorophenol indophenol dye solution. Meanwhile, the amounts of vitamin B1, B2 and B9 were colorimetrically determined as described by Poornima and Ravishankar [16], Uraku *et al.*, [17] and AOAC [11]; respectively.

Evaluation of the Medicinal Efficacy of Fenugreek Seedlings

To study the medicinal efficiency of the seedlings, water and methanolic extracts were prepared to determine their antioxidant and antimicrobial activities. In addition, the amount of some medically-active phenolics was recorded in those extracts.

Preparation of water and methanolic plant extracts:

To obtain water extracts, known weight of the tissues dried at 50°C was incubated in distilled water at 70°C for 20 minutes then filtered. Meanwhile, the dry plant tissues were shaken in methanol at 37°C for 3 hours followed by filtration to obtain the methanolic extracts.

Determination of antioxidant activity:

The procedure given by Ebrahimzadeh *et al.*, [18] was adopted to demonstrate 2, 2'-diphenyl-1-picrylhydrazyl (DPPH)- scavenging capacity of the plant extracts. The plant tissue (in mg) required to reduce initial DPPH concentration by 50% (IC₅₀) was computed graphically. Moreover, the antiradical power (ARP) of the extracts was calculated as the reciprocal of IC₅₀. To explore 2, 2'-azino-bis 3-ethyl benzothiazoline-6-sulfonic acid (ABTS)- scavenging capability of the extracts, the method of Re *et al.*, [19] was followed. The extracts were also assessed for their H₂O₂-scavenging ability following the spectrophotometric protocol of Keser *et al.*, [20]. In addition, the reducing power was investigated following Yildirim *et al.*, [21].

Determination of antimicrobial activity:

Antimicrobial assay was performed via filter paper disc method described by Murray et al., [22]. Stock cultures of Klebsiella pneumonia, Escherichia coli, Erwinia carotovora, Bacillus subtilis, Proteus vulgaris, Streptococcus pyrogenes, Staphylococcus epidermidis, Enterobacter cloacae, Pseudomonas aerugenosa and Candida albicans were obtained from Laboratory of Microbiology, Faculty of Medicine, Mansoura University, Egypt. Nutrient agar medium for bacteria as well as Czapek dox agar for the fungus was used. Ampicillin as an antibacterial agent and clotrimazole as an antifungal drug were employed as positive controls. The microbial susceptibility was evaluated by measuring the inhibition zone diameter.

Determination of phenolic compounds:

Following the spectrophotometric analyses cited from Lin and Tang [23] and Sadasivam and Manickam [24], Folin-Ciocalteu reagent and vanillin hydrochloride reagent were used to record the amount of total phenols and tannins; respectively. In addition, gravimetric methods were adopted to determine the amount of saponins [25], flavonoids [26] and alkaloids [27].

Results

Changes in Nutritional Value of Fenugreek Seedlings

In comparison with the unsalted fenugreek seedlings, application of NaCl at 50 and 200 ppm increased moisture content of the 2- day old seedlings; while 100, 150 and 200 ppm NaCl increased their moisture content when they became 5- day old. Otherwise, a drop-in moisture content of the studied seedlings was recorded. During the two stages, all salt concentrations could raise protein and nitrogen content of the seedlings with corresponding reduction in the amount of reducing, non-reducing and total sugars (Figure 1).



Figure 1: Effect of different salt concentrations on chemical composition (%) of fenugreek seedlings at two different germination stages.

With respect to heavy metals, the amount of Fe generally decreased by salt application at the two addressed stages. In the 2- day old seedlings, Zn and Cu amounts increased by almost all salt levels; while these metals increased by some salt levels when the seedlings were 5- day old (Zn by 100 and 150 ppm and Cu by 50, 100 and 200 ppm). Except under these levels, a decline in the two metals in the 5- day old seedlings was noticed (Figure 2).

Regarding the assayed vitamins, vitamin C increased by most the applied salt concentrations (100, 150 and 200 ppm) in the first stage and the converse was observed in the second stage. For vitamin B1, B2 and B9; they were increased by almost all salt concentrations in the second stage but decreased in the first one because of salt (Figure 3).

Changes in Medicinal Efficacy of Fenugreek Seedlings

Away from DPPH- scavenging activity, the methanolic extracts of fenugreek seedlings generally possessed higher antioxidant capacity than water extracts. Both of water and methanolic extracts of the 2- and 5- day old



Figure 2: Effect of different salt concentrations on heavy metals contents (mg 100 g⁻¹) of fenugreek seedlings at two different germination stages.



Figure 3: Effect of different salt concentrations on vitamins contents (mg g⁻¹) of fenugreek seedlings at two different germination stages.

seedlings had enhanced H_2O_2 - scavenging activity when treated with salt at different concentrations. For the reducing power as well as ABTS- and DPPHscavenging activities of the extracts, these were decreased in the 2- day old seedlings but increased in the 5- day old ones grown under stress (Figure 4).



Figure 4: Effect of different salt concentrations on antioxidant activities of fenugreek seedlings at two different germination stages.

Figure 5: Effect of different salt concentrations on phenolics contents (%) of fenugreek seedlings at two different germination stages.

Concerning the antimicrobial activity represented in table 1, the suppressing effect of water extracts was enhanced by some salt concentrations particularly against Klebsiella pneumonia (by 50, 100 and 150 ppm NaCl in the second stage), Erwinia carotovora (by 50, 100 and 150 ppm in the first stage), Bacillus subtilis (by 100 ppm in the first stage), Candida albicans (by 50 ppm in the second stage) as well as Streptococcus pyrogenes and Staphylococcus epidermidis (both by 50, 100 and 200 ppm in the first stage in addition to 100 and 200 ppm in the second stage). For methanolic extracts, some salt concentrations could enhance the antimicrobial potentiality against Erwinia carotovora (200 ppm in the first stage), Bacillus subtilis (50 ppm in the first stage) and Candida albicans (150 ppm in the second stage).

Escherichia coli, *Proteus vulgaris*, *Enterobacter cloacae* and *Pseudomonas aerugenosa* all seemed to be resistant to the extracts of both stressed and unstressed fenugreek seedlings so these were not represented in table 1.

As observed for the antioxidant activity, methanolic extracts of fenugreek seedlings generally contained higher amount of phenolic compounds than water extracts. In the first stage, tannins in water extracts were the only class of phenolics that increased when applying NaCl particularly at 50 and 100 ppm. In the second stage and in both water and methanolic extracts; tannins, alkaloids, flavonoids and total phenols were all increased by 50 and 150 ppm NaCl solution while saponins increased by 50, 100 and 150 ppm (Figure 5).

Table 1: Effect of different salt concentrations on the antimicrobial activity of fenugreek seedlings at two different germination stages. WE = water extract; ME = methanolic extract; Nt = not tested.

| | | Inhibition Zone Diameter (mm) | | | | | | | | | | | |
|-------------------------|-------------------------------|-------------------------------|----|-----------------------|----|----------------------|----|----------------------------|----|-------------------------------|----|---------------------|----|
| Germination Stage | Salt Concentration (mM) | Klebsiella pneumonia | | Erwinia carotovora | | Bacillus subtilis | | Streptococcus pyrogenes | | Staphylococcus epidermidis | | Candida albicans | |
| | | WE | ME | WE | ME | WE | ME | WE | ME | WE | ME | WE | ME |
| | 0 | 0 | 0 | 0 | 0 | 9 | 0 | 7 | 0 | 7 | 0 | 7 | 7 |
| 2- Day Old | 50 | 0 | 0 | 7 | 0 | 7 | 8 | 10 | 0 | 8 | 0 | 0 | 0 |
| Seedlings | 100 | 0 | 0 | 7 | 0 | 10 | 0 | 8 | 0 | 9 | 0 | 0 | 0 |
| | 150 | 0 | 0 | 7 | 0 | 0 | 0 | 0 | 0 | 7 | 0 | 0 | 6 |
| | 200 | 0 | 0 | 0 | 7 | 7 | 0 | 9 | 0 | 8 | 0 | 0 | 0 |
| 5- Day Old Seedlings | 0 | 7 | 0 | 0 | 0 | 8 | 0 | 7 | 0 | 0 | 0 | 7 | 0 |
| | 50 | 8 | 0 | 0 | 0 | 7 | 0 | 8 | 0 | 8 | 0 | 8 | 0 |
| | 100 | 8 | 0 | 0 | 0 | 7 | 0 | 8 | 0 | 8 | 0 | 0 | 0 |
| | 150 | 8 | 0 | 0 | 0 | 0 | 0 | 8 | 0 | 8 | 0 | 7 | 8 |
| | 200 | 0 | 0 | 0 | 0 | 0 | 0 | 8 | 0 | 8 | 0 | 7 | 0 |
| Antibiotic Drugs | | 27 | | Nt | | Nt | | 25 | | Nt | | 15 | |

Discussion

Plants can synthesize vast array of chemicals in the form of primary or secondary metabolites; and these phytochemicals can be utilized by human as valuable nutrients and to cure or prevent various ailments. *Via* the present study, the impact of salt stress on phytonutrients of fenugreek seedlings was addressed. The moisture content of the studied 2- and 5- day old seedlings increased under most of the applied NaCl concentrations. A similar increase in moisture content of fenugreek plants at successive growth stages under salt treatment was lately recorded by Kapoor and Pande [28] who ascribed that pattern of change in moisture content to the deteriorative status of the plant cells under stress.

Concerning total protein and nitrogen contents of the concerned fenugreek seedlings, these were increased in response to the application of all the used salt concentrations. Matching our findings, Kumar *et al.*, [29] recorded an increase in protein content of some rice cultivars as a result of increasing salinity level up to 100 mM NaCl. Also, Abdul Qados [30] reported a comparable increase in protein content of bean plants with raising NaCl concentrations till 240 mM. Such an alteration in the amount of protein and nitrogen may be an adaptive response to salt stress since some plants can

resist osmotic stress by accumulating certain osmolytes such as soluble proteins and other nitrogenous compounds that enable their cells to absorb more water. Coinciding with this assumption, Kosová *et al.*, [31] pointed out that an active salinity acclimation is conferred by stimulated biosynthesis of certain novel proteins with osmoprotective functions.

With respect to sugar content, a reverse trend was observed herein where salt stress decreased the amount of reducing, non- reducing and total sugars in fenugreek seedlings. Similarly, and at 12 dS m^{-1} salinity level, the amount of reducing, non- reducing and total sugars in the tissues of five Malaysian rice varieties decreased in comparison with their unstressed equivalents [32]. In the current investigation, the decline noticed in seedling sugar content under salt stress may result from lower ability of the stressed seedlings to synthesize sugars and/ or higher rate of sugar degradation to compensate the disturbing conditions [33].

Regarding the amount of heavy metals in the explored fenugreek seedlings, the amount of Fe generally decreased by salt application at the two addressed stages, while Zn and Cu amounts fluctuated among the various treatments. Almost all the studies conducted to assess heavy metal composition of certain plants under the effect of salt stress have highlighted the impact of salinity on the plant ability to uptake heavy metal from the growing medium [34, 35]. However, the case in the present study is completely different since the growing medium of fenugreek seedlings herein is simply NaCl solution at different concentrations (0, 50, 100, 150 or 200 ppm). So, the fluctuation in heavy metals concentrations in the addressed seedlings in response to raising salt concentration can be explained only on the basis of the metabolism of metals already present in the seeds before applying NaCl stress.

In the current investigation, the reduction in Fe content of salinized fenugreek seedlings may indicate higher consumption of Fe reserve under stress conditions in a trial to cope with salinity. Fe is an essential micronutrient for plant cells particularly at the juvenile stage when it is involved in various biochemical and physiological pathways. Fe plays an important role in protein and DNA biosynthesis [36]. Furthermore, Fe acts as a component of some vital enzymes such as cytochromes of the respiratory electron transport chain [37]. Fe is also an active cofactor for many enzymes required for synthesis of the plant hormones ethylene and abscisic acid which are active in many plant development pathways and their adaptive responses to the fluctuating environmental conditions [38].

Unlike the case for Fe, the fenugreek seedlings addressed herein contained higher amounts of Zn and Cu when treated by almost all salt levels in the first stage and some salt levels in the second stage (higher Zn by 100 and 150 ppm; and Cu by 50, 100 and 200 ppm). The higher content of such metals in the salt- stressed seedlings may refer to less ability of the seedlings to benefit from the reserve of these elements when stressed. The two elements are essential for plant cells and are involved in various metabolic processes.

For plants, Zn is an essential micronutrient involved in a wide range of physiological processes [39]. Generally, Zn is involved in protein synthesis [40], auxin anabolism [41] and stabilization of cellular membranes [42]. More interestingly and according to Tavallali *et al.*, [43], Zn can reduce the harmful impact of salt stress where Zn can affect the plant capacity for both water uptake and transport. In cellular membranes, some ligands such as cysteine and histidine can bind to Zn minimizing the production of toxic hydroxyl radicals [41]. Furthermore, there are some evidences that Zn is involved in oxidative stress- induced expression of genes encoding antioxidant defense enzymes such as ascorbate peroxidase and glutathione reductase [44].

With respect to Cu, it is also one of the essential micronutrients required for growth and development of plants. *In vivo*, Cu can exist in different oxidation forms; mainly Cu²⁺ and Cu⁺. According to Raven *et al.*, [45], Cu is a redox- active metal that serves as a structural element in various regulatory proteins. In addition, Cu participates in cell wall metabolism, hormonal signaling

as well as respiratory reactions. Furthermore, Cu plays an important role in oxidative stress responses. Cu ions also act as cofactors for many essential enzymes such as amino oxidase, cytochrome c oxidase, superoxide dismutase and polyphenol oxidase. Moreover, Cu is vital for iron mobilization [46].

In addition to moisture content as well as the amount of protein, nitrogen, sugars and heavy metals investigated in fenugreek tissues in the present study, some vitamins were also quantified under the experimental conditions. The amount of vitamin c in the seedlings generally increased by salt application in first stage but decreased in the second stage. The reverse was recorded for vitamin B1, B2 and B9 that generally increased in the second stage because of salt treatments but decreased in the first stage. A wide range of observations was reported concerning the alteration in vitamins content in plants exposed to salinity. However, it is well documented that some plants suffering different forms of stressful conditions tend to over- synthesize certain water soluble vitamins (vitamin C and some vitamins of B group) with potent antioxidant activity [47].

In accordance with our finding in the first stage, increased ascorbic acid (vitamin c) content in response to salt treatment was previously recorded in Hordeum vulgare [48], Lycopersicon esculentum [49] and Cicer arietinum [50]. Accumulation of ascorbic acid in stressed plants may be an adaptive strategy to cope with the unsuitable conditions. According to Asensi-Fabado and Munné-Bosch [51], ascorbic acid can scavenge reactive oxygen species (ROS), particularly singlet oxygen, preventing the oxidative damage of essential macromolecules (nucleic acids, lipids and proteins) in cytosol, chloroplasts, mitochondria and peroxisomes. However, lowered ascorbate content in the elder fenugreek seedling (5 days old) reported herein in response to salinity may reveal lower capability of seedlings to synthesize such vitamin or higher degrading rates.

Ratnakar and Rai [52] reported that thiamine (vitamin B1) content of Atriplex hortensis plants decreased under NaCl treatment at 60 mM compared to the unstressed plants. On the other hand, Tunc-Ozdemir et al., [53] found that salt- stressed Arabidopsis thaliana seedlings had more thiamine content than their unstressed relatives. Also, Rapala-Kozik et al., [54] observed upgraded levels of thiamine content in NaCl- treated Zea mays plants. According to Tunc-Ozdemir et al., [53], stress- modulated increase in thiamine amount may at least in part result from the enhanced expression of genes coding thiamine synthesis. They also documented that thiamine over- production in stressed plants could stimulate their tolerance to salt- induced oxidative stress. In this context, thiamine was proven to have the ability to quench certain types of ROS especially superoxide anions and hydroxyl radicals [53]. Moreover, thiamine was suggested to have an indirect role in the antioxidative defense system through the provision of more NAD(P)H under stressful conditions [47].

In the leaves of 20 mM NaCl- treated Atriplex hortensis plants, riboflavin (vitamin B2) content remained as the same as that in control plants but it was significantly decreased at 40 and 60 mM NaCl concentrations [52]. On contrary, Mickky et al., [9] found that riboflavin amount in Medicago sativa plants was significantly increased in response to water stress. Although riboflavin was not recognized as a direct antioxidant, some studies have indicated that this vitamin has the ability to induce the accumulation of potent antioxidants more obviously in plants grown under stress [55]. In addition, some riboflavin derivatives, especially FAD, were found to be needed for the activity of many antioxidant enzymes involved in H2O2 scavenging [56]. Moreover, riboflavin was recently recognized as an important regulator of the cellular redox status and is hence involved in the complex network forming the antioxidant defense system [47].

To the best of our knowledge, studies on the stressinduced changes in folic acid (vitamin B9) content of higher plants are to somewhat scarce. However, the common trend in the past few years is directed to exogenous application of folate [57] or folate biofabrication through metabolic engineering [58] to induce potent effects on plant response to stress. Generally, folate and its derivatives have various metabolic functions the most critical of which is the biosynthesis of amino acids, nucleic acids and pantothenate. Moreover, this vitamin is essential for lignin formation and is involved also in photorespiration [59]. Like riboflavin, folate was also recognized as a regulator of the cellular redox state [47].

The antioxidant activity of fenugreek seeds, germinated seeds as well as the vegetative parts is well documented using various antioxidant assays. A study by Dixit et al., [60] has revealed potent antioxidant activity of germinated fenugreek seeds indicated by ferric reducing power, scavenging of DPPH and ABTS radicals, oxygen radical absorbance capacity, pulse radiolysis and inhibition of lipid peroxidation. In that study, the authors ascribed fenugreek antioxidative activity at least partly to the presence of flavonoids and polyphenols. In addition, Kaviarasan et al., [61] recorded that fenugreek seed extract could scavenge DPPH, ABTS as well as hydroxyl radicals and inhibit H2O2- induced lipid peroxidation. They similarly assigned the potency of fenugreek extracts to protect cellular structures from oxidative damage to their high phenolic contents. More recently, methanolic extract of fenugreek seeds were assessed for their antioxidant properties by Pathak et al., [62] who proved that such extract exhibited significant potentiality for iron reduction as well as the scavenging of hydroxyl and DPPH radicals. They also attributed these activities to fenugreek phenolic contents.

Nevertheless and to our best knowledge, almost no attention is paid to study the antioxidant activity of fenugreek grown or germinated under stressful conditions. In the current research, both aqueous and methanolic extracts of fenugreek seedling were evaluated for their *in vitro* antioxidative activities. In most cases, H_2O_2 - scavenging activity of water and methanolic extracts of 2- and 5- day old seedlings were enhanced under the treatment of almost all the tested salt doses. However, the reducing power as well as ABTS- and DPPH- scavenging activities of the extracts were enhanced by salt application only in the 5- day old seedlings; and these were suppressed by salt when the seedlings were younger (2 days old).

In accordance with our results, Chunthaburee et al., [63] found that salt stress could promote the antioxidant activities, evaluated by ferric reducing power as well as DPPH- and ABTS- scavenging activity, in the grains of four Oryza sativa cultivars. In a similar trend, the antioxidant potential of three plants, Salsola baryosma, Trianthema triquetra and Zygophyllum simplex, was found to be elevated under salt treatment as indicated by DPPHscavenging activity and the total reducing power [64]. Furthermore, Abbas et al., [65] reported an enhancement in DPPH- and H₂O₂- scavenging activities as well as total antioxidant capacity and reducing power of Medicago sativa plants suffering water deficit. In 2month old rosemary plants subjected to three salt concentrations (50, 100, and 150 mM NaCl), DPPHscavenging ability of the plant methanolic extracts was enhanced as compared with the unstressed plants [66].

Concerning the antimicrobial activity of fenugreek seedlings studied herein, the suppressing effect of their water extracts were enhanced by some salt concentrations particularly against *Klebsiella pneumonia*, *Erwinia carotovora*, *Bacillus subtilis*, *Candida albicans*, *Streptococcus pyrogenes* and *Staphylococcus epidermidis*. For methanolic extracts, some salt concentrations could enhance their antimicrobial potentiality against *Erwinia carotovora*, *Bacillus subtilis* and *Candida albicans*.

The antibacterial and antifungal activity of different parts of fenugreek plant, especially seeds and germinated seeds, was previously established in many reports. Walli et al., [67] proved potent antimicrobial activity of boiling water extract of unpowdered fenugreek against three Staphylococcus genera namely S. aureus, S. epidermis and S. saprophyticus. El Nour et al., [68] also recorded that petroleum ether extract of fenugreek seeds showed marked antimicrobial activity against Escherichia coli, Staphylococcus aureus, Aspergillus niger and Candida albicans. Moreover, the methanolic extracts of fenugreek calli derived from hypocotyls and cotyledons showed antimicrobial activities against Staphylococcus aureus and Escherichia coli compared to methanolic extracts of seeds that showed antifungal activity against Candida albicans.

According to Barnes *et al.*, [69], fenugreek seeds are rich with the polysaccharide galactomannan in addition to disogenin, yamogenin, gitogenin, tigogenin, neotigogens, choline and trigonelline; all of which are bioactive constituents that could explain the antimicrobial potency of fenugreek. More importantly, the phenolic skeleton of fenugreek may contribute to

their antimicrobial and antioxidant efficiency [70]. Therefore, it was of special significance in the current investigation to study the pattern of change in the amount of total phenols and the main phenolic classes as well in response to the applied NaCl stress.

With respect to phenolics estimated in the present study, tannins in the water extracts of fenugreek seedlings were the only class of phenolics that increased when applying NaCl (50 and 100 ppm) in the first stage. In the second stage and in both water and methanolic extracts; tannins, alkaloids, flavonoids and total phenols were increased when applying 50 and 150 ppm salt solutions while saponins increased by 50, 100 and 150 ppm NaCl in the concerned fenugreek seedlings. Coinciding with the results obtained herein and working also on fenugreek, Hussein and Aqlan [71] found that 0.1% NaCl solution could increase the total amount of phenolics, flavonoids and tannins as well. Meanwhile, higher NaCl concentrations (0.3%) decreased the amount of such metabolites in comparison with the control plants. In a more or less similar manner, Abd Elhamid et al., [72] as well as Sadak [73] recently recorded that water stress caused marked increase in the total phenolic contents as well as total flavonoid and tannin contents in fenugreek plants as compared with their unstressed correspondings. In other plants, various studies have revealed marked overproduction of phenolic compounds in response to stressful conditions. Among those, data recorded by Mickky et al., [9] revealed that application of water stress caused marked increase in the amount of total phenols, saponins, tannins, flavonoids and alkaloids of alfalfa plants; and the increase in the amount of those phenolics was directly proportional to the level of water stress.

Within the plant tissues, phenolics are well documented as stress induced metabolites over- produced in response to various stress stimuli. The accumulation of phenolics under unfavorable growth conditions may be a mechanism exerted by the stressed plants to withstand stress by participating in ROS scavenging mainly through the antioxidative enzymes utilizing polyphenols as co-substrates [74]. In a series of experimental observations, the increase in the titer of total phenols, saponins, tannins, flavonoids and alkaloids in plants growing in a stressing environment was attributed to the enhanced activity of the enzymes involved in phenols biosynthesis and/ or the inhibited activity of the enzymes responsible for their degradation [75, 76].

Conclusion

Based on the results obtained herein, it can be inferred that salinizing fenugreek seedlings can be simply employed to enhance their nutritional value by increasing the amount of some vital phytonutrients such as proteins and certain vitamins. In addition, such strategy can also improve the medicinal value of fenugreek seedlings by upgrading their antimicrobial and antioxidant activities as well as increasing the amount of some medically- active phenolics.

References

- Moradi kor N, Didarshetaban MB, and Saeid Pour HR. "Fenugreek (*Trigonella foenum-graecum* L.) as a valuable medicinal plant." *International journal of Advanced Biological* and Biomedical Research 1 (2013): 922-931.
- Lust JB. "The Herb Book." Bantam Books Inc., New York (1986): 1-55.
- 3. Sadeghzadeh-Ahari D, Kashi AK, Hassandokht MR, Amri A, and Alizadeh Kh. "Assessment of drought tolerance in Iranian fenugreek landraces." *Journal of Food, Agriculture and Environment* 7(2009): 414-419.
- Duke AJ. "Handbook of Legumes of World Economic Importance." Plemus Press, New York and London (1986): 345-355.
- Aghaei K, and Komatsu S. "Crop and medicinal plants proteomics in response to salt stress." *Frontiers in Plant Science* 31 (2013): 4:8.
- Taïbi K, Taïbi F, Abderrahim LA, Ennajah A, Belkhodja M, and Mulet JM. "Effect of salt stress on growth, chlorophyll content, lipid peroxidation and antioxidant defence systems in *Phaseolus vulgaris* L." *South African Journal of Botany* 105(2016): 306-312.
- Kiran kumari SP, Sridevi V, and Chandana Lakshmi MVV. "Studies on effect of salt stress on some medicinal plants." *International Journal of Computational Engineering Research* 2 (2012): 143-149.
- Moghbeli E, Fathollahi S, Salari H, Ahmadi G, Saliqehdar F, Safari A, and Grouh MSH. "Effects of salinity stress on growth and yield of *Aloe vera* L." *Journal of Medicinal Plants Research* 6(2012): 3272-3277.
- Mickky BM, Abbas MA, and El-Shhaby OA. "Economic maximization of alfalfa medically- active phytoconstituents." *International Journal of Bioassays* 5 (2016): 4984-4990.
- Mickky BM, Abbas MA, and El-Shhaby OA. "Economic maximization of alfalfa antimicrobial efficacy using stressful factors." *International Journal of Pharmacy and Pharmaceutical sciences* 8 (2016): 299-303.
- AOAC. "Official methods of analysis of AOAC (Association of Official Analytical Chemists) International; 17th edition." Gaithersburg, MD, USA (2000).
- Masuko T, Minami A, Iwasaki N, Majima T, Nishimaru S, and Lee YC. "Carbohydrate analysis by a phenol-sulphuric acid method in microplate format." *Analytical Biochemistry* 339 (2005):69-72.
- Miller GL. "Use of dinitrosalicyclic acid reagent for determination of reducing sugar." *Analytical Chemistry* 31 (1959): 426-428.
- Allen S, Grimshaw HM, Parkinson JA, and Quarmby C. "Chemical Analysis of Ecological Materials." Oxford; Blackwell (1974).
- 15. Ogunlesi M, Okiei W, Ofor E, and Awonuga O. "Determination of the concentrations of zinc and vitamin

C in oysters and some medicinal plants used to correct male factor infertility." *Journal of Natural Products* 2 (2009): 89-97.

- Poornima GN, and Ravishankar RV. "Evaluation of phytonutrients and vitamin contents in a wild yam, *Dioscorea belophylla* (Prain) hains." *African Journal of Biotechnology* 8 (2009): 971-973.
- 17. Uraku AJ, Okala ANC, and Ibiam UA. "Effect of Spilanthes uliginosa (sw), Ocimum basilicum, Hyptis spicigera and Cymbopogon citratus leaf extracts on biochemical and histological parameters of mice exposed to Plasmodium berghei." PhD Thesis submitted to the Department of Biochemistry, Ebony State University, Abakaliki, Nigeria (2014).
- Ebrahimzadeh MA, Nabavi SM, Nabavi SF, and Eslami B. "Free radical scavenging ability of methanolic extract of *Hyosyamus squarrosus* leaves." *Pharmacologyonline* 2 (2009): 796-802.
- Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, and rice-Evans C. "Antioxidant activity applying an improved ABTS radical cation decolorization". *Free Radical Biology and Medicine* 26 (1999): 1231-1237.
- Keser S, Celik S, Turkoglu S, Yilmaz O, and Turkoglu I. "Hydrogen peroxide radical scavenging and total antioxidant activity of hawthorn." *Journal of Chemistry* 2 (2012): 9-12.
- Yildirim A, Mavi A, and Kara A. "Determination of antioxidant and antimicrobial activities of *Rumex crispus* L. extracts." *Agricultural and Food Chemistry* 49 (2001): 4083-4089.
- Murray R, Rosenthal S, Kobayashi S, and Pfaller A. "Medical Microbiology; 3rd edition." St. Louis: Mosby (1998).
- 23. Lin JY, and Tang CY. "Determination of total phenolic and flavonoid contents in selected fruits and vegetables, as well as their stimulatory effects on mouse splenocyte proliferation." *Food Chemistry* 101 (2007): 140-147.
- Sadasivam S, and Manickam A. "Biochemical Methods; 3rd edition." New Delhi, India: New Age International Ltd (2008).
- 25. Obadoni BO, and Ochuko PO. "Phytochemical studies and comparative efficacy of the crude extracts of some homostatic plants in Edo and Delta States of Nigeria." *Global Journal of Pure and Applied Sciences* 8 (2001): 203-208.
- Dewanto V, Wu X, Adom KK, and Liu RH. "Thermal processing enhances the nutritional value of tomatoes by increasing total antioxidant activity." *Agricultural and Food Chemistry* 50 (2002): 3010-3014.
- 27. Harborne JB. "Phytochemical Methods." London: Chapman and Hall Ltd (1973).
- Kapoor N, and Pande V. "Effect of salt stress on growth parameters, moisture content, relative water content and photosynthetic pigments of fenugreek variety RMt-1." *Journal of Plant Sciences* 10 (2015): 210-221.

- Kumar V, Shriram V, Jawali N, and Shitole MG. "Differential response of indica rice genotypes to NaCl stress in relation to physiological and biochemical parameters." *Archives of Agronomy and Soil Science* 53 (2007): 581-592.
- Abdul Qados AMS. "Effect of salt stress on plant growth and metabolism of bean plant *Vicia faba* (L.)." *Journal of the Saudi Society of Agricultural Sciences* 10 (2011): 7-15.
- Kosová K, Prášil IT, and Vítámvás P. "Protein contribution to plant salinity response and tolerance acquisition". *International Journal of Molecular Sciences* 14 (2013): 6757-6789.
- 32. Hakim MA, Juraimi A, Hanafi MM, Ismail MR, Selamat A, Rafii MY, and Latif MA. "Biochemical and anatomical changes and yield reduction in rice (*Oryza sativa* L.) under varied salinity regimes." *Biomed Research International* (2014) doi: 10.1155/2014/208584.
- 33. Prado FE, Boero C, Gallardo M, and Gonzalez JA. "Effect of NaCl on germination, growth and soluble sugar content in *Chenopodium quinoa* Willd. seeds." *Botanical Bulletin of Academia Sinica* 41 (2000): 27-34.
- Leblebici Z, Aksoy A, and Duman F. "Influence of salinity on the growth and heavy metal accumulation capacity of *Spirodela polyrrhiza* (Lemnaceae)." *Turkish Journal* of Biology 35 (2011): 215-220.
- 35. Kadkhodaie A, Kelich S, and Baghbani A. "Effects of salinity levels on heavy metals (Cd, Pb and Ni) absorption by sunflower and sudangrass plants." *Bulletin of Environment, Pharmacology and Life Sciences* 1 (2012): 47-53.
- Bashir K, Ishimaru Y, Shimo H, Nagasaka S, Fujimoto M, and Takanashi H. "The rice mitochondrial iron transporter is essential for plant growth." *Nature Communications* 2 (2011): 322-329.
- 37. Rout GR, and Sahoo S. "Role of iron in plant growth and metabolism." *Reviews in Agricultural Science* 3 (2015): 1-24.
- Siedow JN. "Plant lipoxygenase: structure and function." Annual Review of Plant Physiology and Plant Molecular Biology 42 (1991): 145-188.
- Di Baccio D, Kopriva S, Sebastiani L, and Rennenberg H. "Does glutathione metabolism have a role in the defense of poplar against zinc excess?" *New Phytologist* 167 (2005): 73-80.
- Hänsch R, and Mendel RR. "Physiological functions of mineral micronutrients (Cu, Zn, Mn, Fe, Ni, Mo, B, Cl)." *Current Opinion in Plant Biology* 12 (2009): 259-266.
- Brennan RF. "Zinc application and its availability to plants." Ph. D. dissertation, School of Environmental Science, Division of Science and Engineering, Murdoch University (2005).
- Dang H, Li R, Sun Y, Zhang X, and Li Y. "Absorption, accumulation and distribution of zinc in highly-yielding winter wheat." *Agricultural Sciences in China* 9 (2010): 965-973.

- 43. Tavallali V, Rahemi M, Eshghi S, Kholdebarin B, and Ramezanian A. "Zinc alleviates salt stress and increases antioxidant enzyme activity in the leaves of pistachio (*Pistacia vera* L. 'Badami') seedlings." *Turkish Journal of Agriculture and Forestry* 34 (2010): 349-359.
- Cakmak I. "Possible roles of zinc in protecting plant cells from damage by reactive oxygen species." *New Phytologist* 146 (2000): 185-205.
- 45. Raven JA, Evans MCW, and Korb RE. "The role of trace metals in photosynthetic electron transport in O₂-evolving organisms." *Photosynthesis Research* 60 (1999): 111-149.
- Yruela I. "Copper in plants." Brazilian Journal of Plant Physiology 17 (2005): 145-156.
- 47. Dubey NK. "Plants as a source of natural antioxidants." CAB International, Wallingford, Oxfordshire, OX10 8DE, UK (2015).
- Sarwat MI, and El-Sherif M. "Increasing salt tolerance in some barley genotypes (*Hordeum vulgare*) by using kinetin and benzyladenine." World Journal of Agricultural Sciences 3 (2007): 617-629.
- 49. Kim HJ, Fonseca JM, Kubota C, Kroggel M, and Choi JH. "Quality of fresh-cut tomatoes as affected by salt treatment in irrigation water and post-processing ultraviolet-treatment." *Journal of the Science of Food and Agriculture* 88 (2008): 1969-1974.
- Mishra M, Mishra PK, Kumar U, and Prakash V. "NaCl phytotoxicity induces oxidative stress and response of antioxidant systems in *Cicer arietinum* L. cv. Abrodhi." *Botany Research International* 2 (2009): 74-82.
- Asensi-Fabado MA, and Munné-Bosch S. "Vitamins in plants: occurrence, biosynthesis and antioxidant function." *Trends in Plant Sciences* 15 (2010): 582-592.
- Ratnakar A, and Rai A. "Influence of NaCl salinity on βcarotene, thiamine, riboflavin and ascorbic acid contents in the leaves of *Atriplex hortensis* L. var. Pusa Bathua No. 1." *Journal of Stress Physiology and Biochemistry* 9 (2013): 187-192.
- Tunc-Ozdemir M, Miller G, Song L, Kim J, Sodek A, Koussevitzky S, Misra AN, Mittler R, and Shintani D. "Thiamine confers enhanced tolerance to oxidative stress in *Arabidopsis*." *Plant Physiology* 151 (2009): 421-432.
- Rapala-Kozik M, Kowalska E, and Ostrowska K. "Modulation of thiamine metabolism in Zea mays seedlings under conditions of abiotic stress." Journal of Experimental Botany 59 (2008): 4133-4143.
- 55. Taheri P, and Tarighi S. "Riboflavin induces resistance in rice against *Rhizoctonia solani via* jasmonate-mediated priming of phenylpropanoid pathway." *Journal of Plant Physiology* 167 (2010): 201-208.
- Gill S, and Tuteja N. "Reactive oxygen species and antioxidant mechinery in abiotic stress tolerance in crop plants." *Plant Physiology and Biochemistry* 48 (2010): 909-930.

- 57. Mohamed NEM. "Behaviour of wheat Cv. Masr-1 plants to foliar application of some vitamins." *Nature and Science* 11 (2013): 1-5.
- 58. Blancquaert D, Van Daele J, Storozhenko S, Stove C, Lambert W, and Van Der Straeten D. "Rice folate enhancement through metabolic engineering has an impact on rice seed metabolism, but does not affect the expression of the endogenous folate biosynthesis genes." *Plant Molecular Biology* 83 (2013): 329-349.
- Hanson AD, and Roje S. "One-carbon metabolism in higher plants." *Annual Review of Plant Physiology and Plant Molecular Biology* 52 (2001): 119-137.
- Dixit P, Ghaskadbi S, Mohan H, and Devasagayam TP. "Antioxidant properties of germinated fenugreek seeds." *Phytotherapy Research* 19 (2005): 977-983.
- Kaviarasana S, Naikb GH, Gangabhagirathic R, Anuradhaa CV, Priyadarsini KI. "In vitro studies on antiradical and antioxidant activities of fenugreek (*Trigonella foenum graecum*) seeds." Food Chemistry 103 (2007): 31-37.
- Pathak N, Pant N, Singh J, and Agrawal S. "Antioxidant activity of *Trigonella foenum graecum* L. using various *in vitro* models." *International Journal of Herbal Medicine* 2 (2014): 53-57.
- 63. Chunthaburee S, Sanitchon J, Pattanagul W, and Theerakulpisut P. "Effects of salt stress after late booting stage on yield and antioxidant capacity in pigmented rice grains and alleviation of the salt-induced yield reduction by exogenous spermidine." *Plant Production Sciences* 18 (2015): 32-42.
- 64. Sharma V, and Ramawat K. "Salt stress enhanced antioxidant response in callus of three halophytes (*Salsola baryosma, Trianthema triquetra, Zygophyllum simplex*) of Thar Desert." *Biologia* 69 (2014): 178-185.
- 65. Abbas MA, El-Shehaby OA, and Mickky BM. "Antioxidant capacity of droughted alfalfa plants grown in different soil types." *Mansoura Journal of Environmental Sciences* 42 (2013): 379-391.
- 66. Kiarostami K, Mohseni R, and Saboora A. "Biochemical changes of Rosmarinus officinalis under salt stress." Journal of Stress Physiology and Biochemistry 6 (2010): 114-122.
- Walli RR, Al-Musrati RA, Eshtewi HM, and Sherif FM. "Screening of antimicrobial activity of fenugreek seeds." *Pharmacy and Pharmacology International Journal* 2 (2015), doi: 10.15406/ppij.2015.02.00028
- El Nour MEM, Ali AMA, and Saeed BAE. "Antimicrobial activities and phytochemical screening of callus and seeds extracts of fenugreek (*Trigonella foenum-graecum*)." *International Journal of Current Microbiology and Applied Sciences* 4 (2015): 147-157.
- 69. Barnes J, Anderson LA, and Phillipson JD. "Herbal Medicines: A Guide for Health Care Professionals." 2nd ed, Pharmaceutical Press, London (2002).

- 70. Wani SA, and Kumar P. "Fenugreek: A review on its nutraceutical properties and utilization in various food products." *Journal of the Saudi Society of Agricultural Sciences* In press (2016), doi: 10.1016/j.jssas.2016.01.007
- 71. Hussein EA, and Aqlan EM. "Effect of mannitol and sodium chloride on some total secondary metabolites of fenugreek calli cultured *in vitro*." *Plant Tissue Culture and Biotechnology* 21 (2011): 35-43.
- 72. Abd Elhamid EM, Sadak MS, and Tawfik MM. "Physiological response of fenugreek plant to the application of proline under different water regimes." *Research Journal of Pharmaceutical, Biological and Chemical Sciences* 7 (2016): 580-594.
- 73. Sadak MS. "Mitigation of drought stress on fenugreek plant by foliar application of trehalose." *International Journal* of Chem Tech Research 9 (2016): 147-155.

- 74. Sgherri C, Cosi E, and Navari-Izzo F. "Phenols and antioxidative status of *Raphanus satinus* grown in copper excess." *Physiologia Plantarum* 118 (2003): 21-28.
- 75. Dai LP, Xiong ZT, Huang Y, and Li MJ. "Cadmiuminduced changes in pigments, total phenolics, and phenylalanine ammonia lyase activity in fronds of *Azolla imbricata*." Journal of Environmental Toxicology 21(2006): 505-512.
- 76. Lee BR, Kim KY, Jung WJ, Avice JC, Ourry A, and Kim TH. "Peroxidases and lignification in relation to the intensity of water-deficit stress in white clover (*Trifolium* repens L.)." Journal of Experimental Botany 58 (2007): 1271-1279.

Cite this article as:

Mickky BM, Abbas MA, and Sameh NM. "Effect of salinization on health benefits of fenugreek seedlings" *International Journal of Bioassays* 6.02 (2017): 5245-5255. **DOI:** http://dx.doi.org/10.21746/ijbio.2017.02.004

Source of support: Nil. Conflict of interest: None Declared