



EFFECT OF AQUEOUS MACERATIVES OF SEED POWDER OF *SYZIGIUM CUMINI* (L) ON THE PROTEIN CONTENTS AND MID GUT ENZYME ACTIVITIES IN THE FIFTH INSTAR LARVAE OF SILK WORM, *BOMBYX MORI* (L) (RACE: PM X CSR₂)

Tejas C Korade, Manali T Patil, Kavita H Nimbalkar

Department of Zoology, Shardabai Pawar Mahila College, Shardanagar Tal, Baramati, Dist. Pune- 413115, India

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Abstract: Different concentrations (5.0 ppm; 10.0 ppm; 20.0 ppm & 50.0 ppm) of the aqueous solution of seed powder of *Syzigium cumini* (L) concentrations was used to treat the leaves of mulberry & fed to the fifth instar larvae of polyvoltine, crossbred silkworm, *Bombyx mori* (L) (Race : PM x CSR₂) for first three days; second day & third day; third day (only). The larvae fed with untreated & water treated leaves were also maintained. Bioassays of proteins (S.P. & T.P.) & enzymes (protease & amylase) were carried out on fifth day through the use of mid gut homogenate. Treating the mulberry leaves with herbal preparations (*Syzigium cumini*) & feeding them to fifth instar larvae was found reflected into significant improvement in the levels of proteins (S.P. & T.P.) & velocities of biochemical reactions catalyzed by protease & amylase. The pattern of increase in soluble proteins & total proteins in the mid gut tissue were 32.147 to 90.074 percent & 5.657 to 39.052 percent respectively. The activities of mid gut protease & amylase were increased by 21.444 to 83.706 percent and 14.54 to 52.257 percent respectively. The nutrient contents of seed powder of *Syzigium cumini* (L) serve to improve the digestibility & exert the influence of efficient metabolism in the fifth instar larvae of silkworm, *Bombyx mori* (L). The *Syzigium* seed powder treatment may gear overall biochemical constituency of silkworm larvae, through the significant improvement in the velocity of mid gut enzyme catalyzed biochemical reactions.

Keywords: Errors, Pre-analytical, Analytical, Post analytical, Total testing process, Quality improvement.

INTRODUCTION

Life of insect herbivores is in the orchestrate progression, which closely interlinked with plant metabolites. The biochemical constituents of plants could have been the factors of growth & metamorphosis for insects (Bowers et al, 1966). The phytophagous insects are able to avoid poor quality food or able to select a high quantitative food from variety available to them. The silkworm, *Bombyx mori* (L) is a monophagous insect, feeding exclusively on the leaves of mulberry *Morus alba* (L). It is therefore, essential to improve either food quality or appetite (or both) of larval instars of silkworm for better performance in silk production. The factors responsible to influence the growth, development & subsequent physiology of insect body include: nutritional qualities of food, biochemical status of nutrients in the food, hormonal level in the body & environmental conditions (Murugan & Georgr, 1992). Elements of the insect body are primarily derived from the food source. For silkworm, the leaves of mulberry contain many stimulants (Ito, 1960, 1961; Nayar & Fraenkel, 1962; Ito, et al, 1964; Ito & Hyashiya, 1965). Nutrition quality in silkworm, *Bombyx mori* (L) serves to accelerate the growth, metamorphosis & forms the physiological foundation for sericulture. The leaves of mulberry are the sole source of food for larval instars of silkworm, *Bombyx mori* (L), biochemically constituted with

proteins, lipids, carbohydrates (Murali, 1992) & minerals (Subramanyam Reddy, 1992). Therefore, corresponding diversity of enzymes capable of hydrolyzing the bio-compounds of mulberry is exhibited by mid gut of larval instars of silkworm, *Bombyx mori* (L). The body tissues of larval instars of silkworm, *Bombyx mori* (L) especially, the fat bodies accumulates large quantity of proteins, lipids & glycogen during the development, which is nothing but the reflection of efficient consumption & utilization of nutrient bio-compounds of mulberry leaves. The variation in the food consumption in phytophagous insects may be for varied biochemical processes, ultimately for successful adaptations (Slansky, 1982). It has been suggested that, there is a functional difference between the activity of digestion by the digestive fluid in mid gut & tissue of mid gut. It has been reported by Horie, et al (1963) that, molecular proteins are hydrolyzed into peptides by digestive fluid content & into amino acids with peptidases in the mid gut tissue. Likewise, the polysaccharides are digested in the insect gut lumen by digestive fluid & disaccharides and/or trisaccharides get hydrolyzed into their constituent mono saccharide sugars mainly in the gut tissue (Horie, 1967).

Lipase, the lipid digesting enzyme of the insect mid

*Corresponding Author:

Dr. Tejas C Korade,
Department of Zoology, Shardabai Pawar Mahila College,
Shardanagar Tal, Baramati District,
Pune-413115, India



gut has been reported to have analogy with pancreatic lipase of vertebrates (Yamafuji & Yonezawa, 1935). The efforts towards the qualitative silk production through the improvement in the efficiency of consumption & utilization of food by larval instars of silkworm, *Bombyx mori* (L) include: improvement in the quality of mulberry leaves & supplementation of nutrient bio-compounds like soya protein; potassium iodide, copper sulphate, other mineral salts, herbal products (or drugs) like digoxin (Vitthalrao & Kulkarni, 2011) & kho-go (Desai, et al, 2011). For the purpose to screen correct dosage & appropriate time of feeding mulberry leaves treated with aqueous solution of seeds of *Syzgium cumini* (L), the present study has been planned.

Table.1: Schedule of treating the mulberry leaves with aqueous solution of *Syzgium cumini* (L) & feeding to the fifth instar larvae of silkworm, *Bombyx mori* (L) (Race: PM x CSR₂)

Group	Day for feeding concentration of kho- go (ppm)	↓ 1 →		
		2	3	
o-0	Untreated control	-	-	-
o-I	Water treated control	✓	✓	✓
o-II	Water treated control	-	✓	✓
o-III	Water treated control	-	-	✓
A-1	5 ppm	+	+	+
A-2	5 ppm	-	+	+
A-3	5 ppm	-	-	+
B-1	10 ppm	+	+	+
B-2	10 ppm	-	+	+
B-3	10 ppm	-	-	+
C-1	20 ppm	+	+	+
C-2	20 ppm	-	+	+
C-3	20 ppm	-	-	+
D-1	50 ppm	+	+	+
D-2	50 ppm	-	+	+
D-3	50 ppm	-	-	+

- = Untreated mulberry leaves.
 ✓ = Water treated mulberry leaves
 + = Kho-go treated mulberry leaves.

MATERIAL & METHODS

The disease free layings (DFL) of multi voltine, crossbreed race: PM x CSR₂ of silkworm, *Bombyx mori* (L) were procured through the sericulture unit of Agriculture Development Trust, Malegaon (Baramati). They were processed for incubation (through black boxing); transfer of hatched larvae on the rearing bed of mulberry leaves and reared through the methods prescribed by Krishnaswami, et al (1978) & explained by Khyade (2004). Soon after the fourth moult, the fifth instar larvae were divided into various groups like untreated control(1), water treated control (3) & the treated (12), each with 100 individuals. Four feedings were followed (6 am, 11 am, 4 pm & 10 pm). 100grams of fresh mulberry leaves were used to feed the group of 100 larvae, for each feeding. The seeds of *Syzgium cumini* (L) were collected from the Malegaon Farm of Agriculture Development Trust. They were identified

through the experts at Department of Botany, Shardabai Pawar Mahila College, Shardanagar. The seeds were shade dried and powdered using the domestic mixture. The seed powder (1000 mg) was macerated through the use of distilled water (1000 ml). The macerate was filtered through muslin cloth. The filtrate was serially diluted to prepare aqueous solution of desired strengths. The stock solutions of seed powder were of strength: 05 ppm; 10 ppm; 20 ppm & 50 ppm. The stock solutions of seed powder were prepared freshly before the feeding. 400 ml of aqueous solution of seed powder was used to soak 100 grams of fresh mulberry leaves. The soaking was carried out for half an hour before feeding. The soaked/ treated mulberry leaves were drained off completely & then fed to the fifth instar larvae of silkworm, *Bombyx mori* (L). Hundred grams of mulberry leaves were utilized for feeding each time, for the group of hundred larvae. The untreated control group of larvae was supplied with untreated leaves of mulberry. Water treated group of larvae were supplied with water treated leaves of mulberry. For each concentration (ppm) of seed powder, three groups of larvae were made, out of which the first group was fed with treated leaves of mulberry for the first three days; second group for second day & third day. Third group of larvae for each concentration was supplied with treated leaves for only third day (Table-1). For remaining days, the larvae were fed with untreated leaves of mulberry. The bioassays of activity of mid gut protease & amylase was carried out on fifth day of fifth instar. Twenty larvae from each group were selected randomly; anaesthetized with chloroform soaked cotton pads & dissected for mid gut in chilled saline (0.9 percent NaCl). The larvae were opened from dorsal side; the entire alimentary canal was removed from each larva; flushed with ice cold saline so as to remove the debris of mulberry leaf & washed with ice cold saline.

The mid gut from alimentary canal was separated; washed with saline; blotted & weighed accurately on electronic balance. The mid gut tissue was fragmented & then homogenized in chilled saline. Homogenate was centrifuged at 40°C for 15 min. at 10000 rpm. The supernatant was equalized to the volume, aliquots of which contains 10 mg per ml & used as assay sample. Half the volume of assay sample was utilized for bioassay of soluble proteins & another half for mid gut enzymes (protease & amylase).

Bioassay of soluble proteins was carried out through the methods of Lowery, et al, (1951). For each assay sample (of each group), bioassay was carried in the triplicate set. One ml of assay sample was added in each test tube. The blank test tube was also prepared simultaneously, in which the assay sample was replaced with distilled water. Addition of 5 ml Lowery's

“C” solution was made in each test tube, mixed well & kept for 15 minutes for the purpose to form the copper-protein complex. After fifteen minutes; 0.5 ml Folin’s phenol reagent was added in each test tube & mixed well. The content in each test tube was allowed to develop color. Then the optical density of content of each test tube was recorded at 660 nm on spectrophotometer. The concentration of soluble proteins of each assay sample was calculated through the reference of optical density assay sample & standard proteins (BSA) (the plot of optical density against concentration Of BSA). The experimentations were repeated for thrice to obtain consistent results.

The content of soluble proteins in each assay sample was expressed in the unit as microgram protein per mg tissue.

For the purpose to determine total protein contents of tissue, another set of twenty larvae was selected randomly from each group. They were anaesthetized with chloroform soaked cotton pads & dissected for mid gut tissue.

The mid gut tissue was homogenized in chilled distilled water by using clean & sterilized mortar & pestle in one normal (1.0N) solution of sodium hydroxide & kept at 37°C for 24 hours. Then it was precipitated with equal volume of ten percent solution of TCA & centrifuged at 10000 rpm for 10 minutes. The precipitate was dissolved in 1.0 NaOH & used as assay sample for total proteins. Further methods of determination of contents of total proteins are similar as described for soluble proteins.

The activity of mid gut protease was carried out according to the method of Brik, et al, (1962) with modifications suggested by Isshaya, et al, (1971) & outlined by Chougale (1992) & Khyade (2004). The mid gut protease activity was determined in triplicate set along with the blank. The mixture of incubation consisted of substrate (one ml of ten percent caein solution); source of enzyme (0.5 ml assay sample) & 0.5 ml of 0.2M Trisbuffer (pH= 8.4). For the blank, assay sample was replaced by distilled water. The incubation was carried out in water bath at 30°C for 20 minutes with constant shaking. Addition of 6 ml of 2 percent trichloroacetic acid was made. The content was centrifuged at 8000 rpm for 15 minutes. The

supernatant was used to read the optical density at 280 nm on spectrophotometer. Amount of tyrosine liberated from the casein due to action of mid gut protease was calculated through the use of optical density readings for assay sample; tyrosine (from standard graph) & predetermined soluble protein contents of each assay sample. The activity of mid gut protease was expressed in terms of specific activity: microgram tyrosine liberated per mg protein per minute.

The activity of mid gut amylase was determined according to the methods of Bernfeld (1955); explained by Ishaaya & Swirski (1970), with modifications suggested by Gaikwad (1998) & outlined by Khyade (2004) & Desai, et al, (2011).

For the purpose to determine the activity of mid gut amylase, 20 larvae were selected randomly & processed for assay sample preparation as described for soluble proteins. Mid gut amylase was determined in triplicate set along with blank. The incubation mixture consisted of one ml of one percent starch solution (as substrate), phosphate buffer (pH=9.2) & 0.5 ml of assay sample. For the blank, assay sample was replaced by distilled water. The process of incubation was carried out in water bath at 30°C for 20 minutes. After incubation the termination of activity of enzyme was made by addition of 2ml DNSA & 2ml distilled water. The contents were heated in boiling water bath exactly for five minutes, cooled immediately & the optical density of content was read at 540 nm on spectrophotometer.

For the purpose to calculate the mid gut amylase activity; the optical density readings for each assay sample; standard solution of maltase (from graph) and soluble proteins were utilized. The enzyme activity was expressed in specific activity: micrograms of maltose liberated per mg protein per minute.

The experimentations were repeated for thrice for the purpose to obtain consistency in the results. The collected data was subjected for statistical analysis (mean, standard deviation, percent change & significance through student t – test) by the methods of Norman & Baily (1955).

Table.2: Contents of proteins & activity of enzymes in the mid gut tissue of the fifth instar larvae of silkworm, *Bombyx mori* (L) fed with the leaves of mulberry, *Morusalba* (L) (M-5: variety) treated with aqueous solution of seed powder of *Syzigium cumuni* (L).

Sr. No.	Moiety	Soluble proteins	Total Proteins	Protease activity	Amylase activity
1.	Group Untreated control (o-o)	138.83 (+ 49.851)	579.43 (+ 126.51)	1.786 (+ 0.154)	3.817 (+ 0.229)
2.	Water treated control (o-I)	134.11 (+ 61.486)	566.14 (+ 159.38)	1.797 (+ 0.468)	3.839 (+ 0.897)
3.	Water treated control (o-II)	136.07 (+ 55.668)	567.89 (+ 143.23)	1.786 (+ 0.521)	3.839 (+ 0.897)
4.	Water treated control (o-III)	136.69 (+ 52.579)	571.51 (+ 139.28)	1.791 (+ 0.815)	3.851 (+ 0.914)
5.	A-1	183.46** (+ 52.409)	612.21* (+ 166.35)	2.169*** (+ 0.043)	4.372* (+ 0.631)
6.	A-2	32.147 185.29** (+ 59.931)	5.657 619.16* (+ 143.78)	21.444 2.236** (+ 0.147)	14.54 4.465* (+ 0.871)
7.	A-3	33.465 185.47** (+ 59.126)	6.856 621.09* (+ 151.71)	25.195 2.274** (+ 0.263)	16.976 4.478* (+ 0.889)
8.	B-1	33.595 219.78** (+ 98.213)	7.185 683.27* (+ 298.68)	27.323 2.313*** (+ 0.279)	17.317 4.559* (+ 0.914)
9.	B-2	98.306 238.01** (+ 77.324)	17.921 758.58* (+ 121.29)	29.507 2.889*** (+ 0.348)	19.439 4.842* (+ 0.929)
10.	B-3	71.439 241.79** (+ 68.763)	30.918 758.61** (+ 189.73)	61.758 2.989*** (+ 0.348)	26.853 4.913** (+ 0.782)
11.	C-1	74.162 242.47** (+ 76.296)	30.923 789.43** (+ 134.18)	67.357 3.026*** (+ 0.312)	28.713 5.483** (+ 0.859)
12.	C-2	74.652 242.51** (+79.012)	36.242 789.59 (+247.26)	69.428 3.083*** (+0.617)	43.646 5.489** (+1.012)
13.	C-3	74.681 242.57** (+ 88.136)	36.270 789.63** (+ 313.13)	72.62 3.087*** (+ 0.983)	43.804 5.588** (+ 1.132)
14.	D-1	74.724 263.23** (+ 82.969)	36.277 805.33* (+ 313.21)	72.844 3.098*** (+ 0.623)	46.397 5.673** (+ 0.863)
15.	D-2	89.605 263.47** (+ 89.712)	38.986 805.39* (+ 211.98)	73.46 3.118*** (+ 0.589)	48.624 5.681** (+ 0.782)
16.	D-3	89.778 263.88** (+ 87.609)	38.996 805.71* (+262.59)	74.58 3.281*** (+ 0.616)	48.834 5.888** (+ 8.823)
		90.074	39.052	83.706	54.257

- Each figure is the mean & three replications.

- Figure in parenthesis with + sign is the standard deviation.

- Figure below parenthesis is percent change.

* : P<0.05
** : P<0.01
*** : P<0.001

RESULTS & DISCUSSION

The results on the biochemical response of the mid gut tissue in the fifth instar larvae of polyvoltine, crossbreed, silkworm, *Bombyx mori* (L) to the aqueous solution of *Syzigium cumuni* (L) seed powder treated leaves of mulberry *Morusalba* (L) (M-5: variety) are summarized in table 2 & presented in fig.

Treating the mulberry leaves with various concentrations of aqueous solutions of syzigium seed powder & feeding them to the fifth instar larvae of

silkworm, *Bombyx mori* (L) for first three days; second & third days & for third day (only) was found variously reflected in the levels of contents of proteins (soluble & total) & activity of enzymes (protease & amylase) in the mid gut tissue homogenate. The soluble proteins of untreated control (o-o) & water treated (for first three days) (o-I) control were found measured 138.83 (+ 49.851) & 134.11(+ 61.486) units respectively. The water treated control groups for second day & third day (o-II)

& for only third day were exhibited 136.07 (+ 55.668) & 136.69 (+52.579) units of soluble proteins in their mid-gut homogenate. The contents of total proteins in the mid gut tissue of fifth instar larvae fed with untreated mulberry leaves were measured 579.43 (+ 126.51) units. The group of larvae fed with water treated mulberry leaves for first three days; second day & third day, only for third day were found 566.14 (+ 159.38); 567.89 (+ 143.19) & 571.51 (+139.28) units of total proteins respectively in their mid-gut tissue homogenate. Treating the mulberry leaves with water may affect either digestibility in the mid gut lumen or absorption of digested matter by epithelial surface of mid gut in fifth instar larvae of silkworm, *Bombyx mori* (L) (Vitthalrao Khyade & Jyoti Kulkarni, 2011 & Desai, et al, 2011). To allow the water treated mulberry leaves for complete draining or shade drying may help for the midgut efficiencies in the fifth instars of silkworm, *Bombyx mori* (L).

The contents of soluble & total proteins in the mid gut tissue homogenate of larvae fed with 5 ppm aqueous solution of syzigium seed powder treated mulberry leaves for first three days; second day & third day & third day (only) were found increased by 32.147 to 33.595 & 5.657 to 7.185 percent respectively. The mid gut protease activity & amylase activity in these groups were found elevated from 21.444 to 27.323 and from 14.54 to 17.317 percent respectively. Increase in the concentration of herbal drug for treating mulberry leaves & feeding as per the schedule was found reflected into significant improvement in the levels of proteins (soluble & total) & the activity of enzymes in the mid gut tissue homogenate. Soluble proteins seem to increase significantly in their level (up to 90 percent) & total proteins (up to 39 percent).

The mid gut protease activity in all the groups of herbal drug treatment was found increased significantly. The levels of significance for the improvement in the mid gut amylase activity was found similar for the groups: 5, 6, 7, 8 & 9. The mid gut amylase activity in the groups: 10-16 was found with higher level of significance.

Increase in the levels of proteins (S.P. & T.P.) in the fifth instar larvae of silkworm, *Bombyx mori* (L) fed with mulberry leaves treated with various concentrations of aqueous solutions of syzigium seed powder may be explained away as due to enhanced break down of contents of mulberry leaves through mid-gut enzymes. Contents of seeds of *Syzigium cumini* (L) improve appetite & digestion. Syzigium seed powder may contain insect juvenoids, which are known to increase the capability of consumption & utilization of food by insects like silkworm. In phytophagous insects, the exogenous compounds through herbal feed mimic the action of natural juvenile hormone,

which enhance the synthesis of poly (A) RNA for major silk protein (Sen, 1988). Most significant response for aqueous solution of seed powder of *Syzigium cumini* (L) treatment in the study seems to be the significant levels of soluble proteins & activities of mid gut protease & mid gut amylase. The soluble proteins contribute in the tissue metabolism through enzymes. According to Applebaum (1985), continuous feeding in insects get reflect into advancement of production of mid gut enzymes, which improve the enzyme efficiencies. Most significant improvement in the protease activity in the treated group of study may be concerned with contents of syzigium seed powder. Individual content treatment may screen out the compounds responsible for improved protease and amylase activities.

The final larval instars of lepidopteran insects have four phases of growth which include: preparatory (first two days); Accumulation phase (third & fifth days); Regression phase (sixth day) & Degeneration phase (day of spinning). The initial preparatory phase is characterized by high rate of DNA synthesis, high rate of digestion, moderate RNA synthesis & low protein synthesis. This phase seems to be juvenile hormone dependent. Accumulation phase, regression phase & degeneration phase are concerned mainly with silk glands. Improvement in the levels of mid gut proteins (S.P. & T.P.); efficiency of mid gut protease & amylase in the group of larvae fed with mulberry leaves treated with aqueous solutions of syzigium powder in the present study seems to be affecting the growth phases of larva. Treating the mulberry leaves with aqueous solution of syzigium seed powder & feeding them to the fifth instar larvae of silkworm, *Bombyx mori* (L) for first three days seems to be significant in comparison with others (2). Feeding treated leaves for first three days possibly availing the herbal nutrients, which affect digestibility of larvae & may contribute phyto-juvenoids or other compounds of growth & development. The study should be extended for screening juvenoid activity of *Syzigium cumini* (L) through the use of the fifth instar larvae of silkworm, *Bombyx mori* (L). Larval parameters & commercial parameters in sericulture, which may fortify the concept & exert applicable influence.

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